# COPEPODA ASSOCIATED WITH THE SCLERACTINIAN CORAL GENUS MONTIPORA IN THE INDO-PACIFIC 

Arthur G. Humes


#### Abstract

Six new species and 2 new genera, Hetairosyna and Tychomyzon, are added to the 14 species of copepods already known to be associated with various species of the hard coral genus Montipora in the Indo-Pacific. The new species include the cyclopoid Euryte bellatula, the asterocherid siphonostomatoids Hetairosyna terpna, Hetairosyna laciniata, Hetairosyna sororia, and Tychomyzon petalum, and the lichomolgid poecilostomatoid Haplomolgus incolumis. New host and locality records for the lichomolgids Haplomolgus montiporae Humes \& Ho, 1968, and Odontomolgus forhani Humes, 1978, are given. A list of all copepods known to be associated with Montipora is provided.


The acroporid scleractinian coral genus Montipora is widespread in shallow waters of the tropical Indo-Pacific Ocean, from Madagascar and the Red Sea to Japan, Australia, Hawaii, and the Marquesas (Veron 1986). Two hundred and eleven nominal species are known, but the number of true species is unknown, with 38 true species reported from Australia (Veron 1986). Copepods are associated with species of Montipora in Madagascar (Humes \& Ho 1968a, 1968b; Humes \& Dojiri 1982, 1983), Mauritius (Stock 1966); India (Sebastian \& Pillai 1974); the Moluccas (Humes 1978, Humes \& Dojiri 1982); northeastern Australia (Humes 1978, 1984; Humes \& Dojiri 1982), New Caledonia (Humes 1978, Humes \& Dojiri 1982); and Hawaii (Humes 1984).

In this work a new species of copepod is reported from Montipora at Enewetak Atoll in the Marshall Islands. Other new species, representing two new genera, are described from Madagascar, the Moluccas, northeastern Australia, and New Caledonia. New host and distribution records are recorded for certain species. The number of species of copepods living with members of this large coral genus is now increased from 14
to 20. A list of the corals and their associated copepods is provided.

## Materials and Methods

The collection of the copepods covered a span of 19 years, from 1963-1982. Since the colonies of Montipora were often too large to be collected whole, fragments were isolated in plastic bags in sea water. In the laboratory, the corals were soaked for several hours in sea water with sufficient ethyl alcohol added to make a solution of approximately $5 \%$. After rinsing the corals, the water was passed through a fine net (120 holes per 2.5 cm , each hole approximately $120 \mu \mathrm{~m}$ square). The copepods were then removed from the sediment retained in the net.

The copepods were measured and dissected in lactic acid, using the wooden slide technique described by Humes \& Gooding (1964). All figures were drawn with the aid of a camera lucida. The letter after the explanation of each figure refers to the scale at which it was drawn. The body length does not include the setae on the caudal rami. The lengths of the segments of the first an-
tennae were measured along their posterior nonsetiferous margins. In the formulas for the armature of legs $1-4$ the Roman numerals indicate spines, the Arabic numerals represent setae.

The abbreviations used are: $\mathrm{R}=$ rostrum, $\mathrm{A}_{1}=$ first antenna, $\mathrm{A}_{2}=$ second antenna, S $=$ siphon, $\mathrm{MD}=$ mandible, $\mathrm{P}=$ paragnath, $\mathrm{MX}_{1}=$ first maxilla, $\mathrm{MX}_{2}=$ second maxilla, $M X P D=$ maxilliped, and $P_{1}=\operatorname{leg} 1$.

Order Cyclopoida Burmeister, 1834
Family Cyclopidae Dana, 1853
Genus Euryte Philippi, 1843
Euryte bellatula, new species
Figs. 1, 2, 3
Type material. - 4 \&, 2 oे from Montipora compressa (Esper), in 2 m , Natsepa, Ambon, Moluccas, $03^{\circ} 37^{\prime} 05^{\prime \prime} \mathrm{S}, 128^{\circ} 17^{\prime} 00^{\prime \prime} \mathrm{E}, 28$ May 1975. Holotype $\%$ (USNM 254456), allotype (USNM 254457), and 3 paratypes ( 2 $\%, 1$ o) (USNM 254458) deposited in the National Museum of Natural History, Smithsonian Institution, Washington, D.C. One female paratype (dissected) in the collection of the author.

Other specimens. -6 o from Montipora lobulata Bernard, in 0.5 m , Rocher à la Voile, Noumea, New Caledonia, $22^{\circ} 18^{\prime} 24^{\prime \prime}$ S, $166^{\circ} 5^{\prime} 50^{\prime \prime} \mathrm{E}, 17$ Jun 1971 ( 1 q dissected); 4 +1 , 1 from Montipora caliculata (Dana), in 1.5 m , Ricaudy Reef, near Noumea, $22^{\circ} 19^{\prime} 05^{\prime \prime} \mathrm{S}, 166^{\circ} 26^{\prime} 28^{\prime \prime} \mathrm{E}, 25$ Jun 1971; 1 ㅇ from Montipora ramosa Bernard, in 2 m , north of Isle Maître, near Noumea, $22^{\circ} 19^{\prime} 30^{\prime \prime}$ S, $166^{\circ} 24^{\prime} 35^{\prime \prime}$ E, 13 Jul 1971; 1 ㅇ, 1 ô from Montipora prolifera Brueggemann, in 3 m , southern shore of Goenoeng Api, Banda Islands, Moluccas, $04^{\circ} 32^{\prime} 05^{\prime \prime} \mathrm{S}$, $129^{\circ} 52^{\prime} 30^{\prime \prime}$ E, 26 Apr 1975 (ô dissected); 2 $\$$ from Montipora verrilli Vaughan, in 3 m , Mermaid Cove, Lizard Island, northeastern Australia, $14^{\circ} 39^{\prime} 50^{\prime \prime} \mathrm{S}, 145^{\circ} 27^{\prime} 00^{\prime \prime} \mathrm{E}, 27$ Oct 1982.

Female. - Body (Fig. 1a) slender and elongate. Length $0.83 \mathrm{~mm}(0.80-0.87 \mathrm{~mm})$ and greatest width 0.31 mm (0.30-0.32
mm ), based on 10 specimens. Greatest dorsoventral thickness 0.22 mm . Epimera of segment bearing leg 1 pointed posteriorly, those of segments bearing legs 2 and 3 subtruncate, and those of segment bearing leg 4 rounded. Ratio of length to width of prosome $1.52: 1$. Ratio of length of prosome to that of urosome 1.25:1.

Segment bearing leg 5 (Fig. 1b) $47 \times 161$ $\mu \mathrm{m}$ in dorsal view, pointed posterolaterally. Genital segment in dorsal view (Fig. 1c) elongate, $107 \mu \mathrm{~m}$ long, $86 \mu \mathrm{~m}$ in greatest width, and $58 \mu \mathrm{~m}$ in least width. Lateral spiniform process (Fig. 1c, d) present on both sides almost midway on segment. Genital areas located dorsolaterally near widest part of segment (Fig. 1c), both areas connected by duct to single midventral pore (Fig. 1d, e). Each area with 2 small hyaline setae approximately $20 \mu \mathrm{~m}$ (Fig. 1d). Three postgenital segments from anterior to posterior $39 \times 55,31 \times 52$, and $44 \times 47 \mu \mathrm{~m}$.

Caudal ramus (Fig. 1f) elongate, unornamented, $73 \times 18 \mu \mathrm{~m}$, ratio 4.06:1. Outer lateral seta short, $15 \mu \mathrm{~m}$, dorsal seta $28 \mu \mathrm{~m}$, both smooth. Outermost terminal seta 52 $\mu \mathrm{m}$, innermost terminal seta $96 \mu \mathrm{~m}$, and 2 long median terminal setae $210 \mu \mathrm{~m}$ (outer) and $340 \mu \mathrm{~m}$ (inner), all with lateral setules.

Body surface without visible ornamentation.

Egg sac (seen on only 1 female) (Fig. 1g) containing 2 eggs $125 \times 125 \mu \mathrm{~m}$ and $146 \times$ $120 \mu \mathrm{~m}$.

Rostrum (Fig. 2a, b) linguiform. First antenna (Fig. 2c) $273 \mu \mathrm{~m}$ long, 21 -segmented. Lengths of segments: 28 ( $53 \mu \mathrm{~m}$ along anterior margin), $20,11,7.5,6.5,6.5,6.5,6.5$, $11,12,13,10,9,9,11,9,9,9,12,12$, and $20 \mu \mathrm{~m}$, respectively. Armature: $8,3,2,2$, $2,2,2,2,2,1,1,1,1,0,1,1,1,1,2,2$, and 8 . All setae smooth.

Second antenna (Fig. 2d) 4-segmented, $160 \mu \mathrm{~m}$ long without setae. Armature: 3, 1, 5 , and 6 , with 4 terminal setae slightly geniculate. Fourth segment with row of inner marginal setules.

Labrum (Fig. 2a, e) with median denticles


Fig. 1. Euryte bellatula, female. a, Dorsal (scale A); b, Urosome, dorsal (B); c, Genital segment, dorsal (C); d, Genital segment, lateral (C); e, Segment bearing leg 5 and genital segment, ventral (C); f, Anal segment and caudal ramus, dorsal (C); g, Egg sac, ventral (D).


Fig. 2. Euryte bellatula, female. a, Rostrum, labrum, mandibles, and paragnaths, ventral (scale B); b, Outline of rostrum, lateral (B); c, First antenna, anterodorsal (E); d, Second antenna, postero-inner (E); e, Labrum, ventral (C); f, Mandible, ventral (C); g, Mandible, flat inner view (C); h, First maxilla, inner (C); i, First maxilla, outer (C); j, Second maxilla, inner (C); k, Maxilliped, inner (C); l, Leg 1 and intercoxal plate, anterior (E).
flanked by few setules. Mandible (Fig. 2a, f, g) $105 \mu \mathrm{~m}$ long with several unequal terminal teeth (in groups of 4 and 5) and 2 unilaterally barbed setae, and having small 1 -segmented palp bearing 3 smooth setae. Paragnath small lobe posterior to mandible (Fig. 2a). First maxilla (Fig. 2h, i) with proximal part (gnathobase) having 6 barbed spines and 2 setae with lateral setules, distal part (palp) forming broad lobe with several terminal denticles and 2 pairs of smooth setae. Second maxilla (Fig. 2j) with first segment (praecoxa) unarmed, second segment (coxa) with 1 seta and distal inward expansion having 2 setae bearing lateral setules and forming forceps, third segment (basis) with 3 setae, and small fourth segment (exopod) with 3 long setae and 1 short seta. Maxilliped (Fig. 2k) 3-segmented, first segment (coxa) with 2 setae, second segment (basis) with 1 seta, third segment (endopod) elongate (possibly 3 -segmented but divisions obscure) and having terminally 2 clawlike setae and 1 straight seta, and 1 seta and row of spinules on inner margin.

Legs 1-4 (Figs. 21, 3a-c) biramous with 3 -segmented rami. Spine and setal formula as follows:

| $\mathrm{P}_{1}$ coxa 0-1 basis 1-I | $\exp \mathrm{I}-1 ; \mathrm{I}-1 ; \mathrm{III}, \mathrm{I}, 4$ |
| :---: | :---: |
|  | enp 0-1; 0-2; I,II,III |
| $\mathrm{P}_{2}$ coxa 0-1 basis 1-0 | $\exp$ I-1; I-1; III,I,IV,1 |
|  | enp 0-1; 0-2; I,II,III |
| $\mathrm{P}_{3}$ coxa 0-1 basis 1-0 | $\exp$ I-1; I-1; III,I,V |
|  | enp 0-1; 0-2; I,II,III |
| $\mathrm{P}_{4}$ coxa 0-1 basis 1-0 | $\exp \mathrm{I}-1 ; \mathrm{I}-1 ; \mathrm{II}, \mathrm{I}, \mathrm{V}$ |
|  | enp 0-1; 0-2; I,II,II |

Leg 1 with basis having inner fringed spine $39 \mu \mathrm{~m}$ long. Rami with outer spines fringed, inner spines narrowly lamellate. Third segment of exopod of leg 2 notable in having formula III,I,IV,1.

Leg 5 (Fig. 1e) ventrally placed and united midventrally by narrow sclerite. First segment $26 \times 15 \mu \mathrm{~m}$ with 1 short dorsal seta. Second segment elongate, $49 \times 13 \mu \mathrm{~m}$, unornamented, with 1 outer lateral fringed spine $52 \mu \mathrm{~m}$, and terminally with 2 fringed
spines $65 \mu \mathrm{~m}$ (outer) and $78 \mu \mathrm{~m}$ (inner) and 1 smooth seta $57 \mu \mathrm{~m}$ between them.

Leg 6 probably represented by 2 setae on genital area (Fig. 1d).

Color of living specimens unknown.
Male. - Body (Fig. 3d) slender. Length $0.59 \mathrm{~mm}(0.55-0.61 \mathrm{~mm})$ and greatest width 0.20 mm ( $0.18-0.22 \mathrm{~mm}$ ), based on $4 \mathrm{spec}-$ imens. Greatest dorsoventral thickness 0.16 mm . Ratio of length to width of prosome 1.89:1. Ratio of length of prosome to that of urosome 1.32:1.

Segment bearing leg 5 (Fig. 3e) $31 \times 67$ $\mu \mathrm{m}$. Genital segment $44 \times 57 \mu \mathrm{~m}$, only little wider than long. Four postgenital segments from anterior to posterior $34 \times 42,30 \times$ $39,24 \times 35$, and $29 \times 34 \mu \mathrm{~m}$.

Caudal ramus resembling that of female but smaller, $44 \times 15 \mu \mathrm{~m}$, ratio 2.93:1.

Body surface unornamented as in female.
Rostrum similar to that of female. First antenna (Fig. 3f) $210 \mu \mathrm{~m}$ long, $15-\mathrm{seg}-$ mented, slightly modified. Lengths of segments: 26 (43 $\mu \mathrm{m}$ along anterior margin), $16,15,5,4,4,9,8,3,23,10,10,21,23$, and $29 \mu \mathrm{~m}$, respectively. Armature: 8, 3, 2, $2+1$ aesthete, $2,2,2,2,2,1,2+1$ aesthete, 2, 1, 3, and $8+1$ aesthete. All setae smooth. Second antenna as in female.

Labrum, mandible, paragnath, first maxilla, second maxilla, maxilliped, and legs 14 like those of female.

Leg 5 (Fig. 3 g ) resembling that of female, similarly placed, second segment $33 \times 12$ $\mu \mathrm{m}$.

Leg 6 (Fig. 3h, i) posteroventral flap on genital segment bearing fringed spine $15 \mu \mathrm{~m}$ and 2 slender setae approximately $11 \mu \mathrm{~m}$.

Spermatophore not seen.
Color unknown.
Etymology. - The specific name bellatu$l a$, Latin meaning pretty or charming, alludes to the svelte appearance of this species.

Remarks. - Vervoort (1964:39) listed six nominal species, one variety and one undetermined species in the genus Euryte, and described one new species, Euryte pseudo-


Fig. 3. Euryte bellatula. Female. a, Leg 2 and intercoxal plate, anterior (scale E); b, Leg 3 and intercoxal plate, anterior (E); c, Leg 4 and intercoxal plate, anterior (E). Male: d, Dorsal (A); e, Urosome, dorsal (E); f, First antenna, dorsal (C); g, Leg 5, lateral (F); h, Genital segment with leg 6, ventral (F); i, Genital segment and first postgenital segment, lateral (F).
robusta. Ummerkutty (1970) listed eight species and one variety. As far as can be determined, no new species have been added in recent years, and the number of congeners remains at eight or nine, depending upon whether varieties are treated as species.

Euryte bellatula differs from all previously described species in the genus. The formula III,I,IV, 1 for the third segment of the exopod of leg 2 distinguishes the new species from Euryte robusta Giesbrecht, 1900 (length of the female $1.20-1.40 \mathrm{~mm}$, Sars 1913), E. curticornis Sars, 1913 (short first antenna, Sars 1913, pl. XIV), and E. pseudorobusta Vervoort, 1964 (short caudal ramus, 2:1, Vervoort 1964), all of which have the formula III,I,V for the third segment of the exopod of leg 2. Euryte brevicauda Sewell, 1949, has a caudal ramus about 1.5:1 (Sewell 1949:35). Euryte similis T. Scott, 1912, has a more robust distal segment in leg 5 of the female, and has the formula I,II,III for the third segment of the endopod of leg 4 (T. Scott 1912, pl. 1, 21, 22). Euryte longicauda Philippi, 1843, has long caudal rami, about 9:1 (Sars 1913, pl. XII).

Order Poecilostomatoida Thorell, 1859
Family Lichomolgidae Kossmann, 1877 Genus Haplomolgus Humes \& Ho, 1968

Haplomolgus incolumis, new species Figs. 4, 5, 6

Type material. -128 ㅇ, 51 o from Montipora caliculata (Dana), in 1.5 m , Ricaudy Reef, near Noumea, New Caledonia, $22^{\circ} 19^{\prime}$ $00^{\prime \prime} \mathrm{S}, 166^{\circ} 27^{\prime} 18^{\prime \prime} \mathrm{E}, 25$ Jun 1971. Holotype $\ddagger$ (USNM 254471), allotype (USNM 254470), and 167 paratypes ( 122 \& , 45 o) (USNM 254469) deposited in the National Museum of Natural History, Smithsonian Institution, Washington, D.C. Remaining paratypes in the collection of the author.

Female. - Body (Fig. 4a) elongate, prosome moderately broad. Length 1.19 mm
$(1.11-1.24 \mathrm{~mm})$ and greatest width 0.43 mm ( $0.41-0.44 \mathrm{~mm}$ ), based on 10 specimens. Greatest dorsoventral thickness 0.35 mm . Epimera of segments bearing legs 1-4 rounded. Segment bearing leg 1 fused with cephalosome. Ratio of length to width of prosome 1.68:1. Ratio of length of prosome to that of urosome 1.26:1.

Segment bearing leg 5 (Fig. 4b) $104 \times 221$ $\mu \mathrm{m}$. Genital segment in dorsal view subquadrate, $138 \times 151 \mu \mathrm{~m}$, little wider than long, in lateral view notched dorsally (Fig. 4c). Genital areas situated dorsolaterally in anterior half of segment and connected by transverse, very broadly $U$-shaped bar. Each area with 2 minute setae approximately $5 \mu \mathrm{~m}$ (Fig. 4c). Three postgenital segments from anterior to posterior $78 \times$ $104,81 \times 99$, and $52 \times 65 \mu \mathrm{~m}$.

Caudal ramus (Fig. 4d) elongate, $117 \times$ $21 \mu \mathrm{~m}$, width taken at middle, ratio 5.57:1. Outer lateral seta $57 \mu \mathrm{~m}$, dorsal seta $73 \mu \mathrm{~m}$, outermost terminal seta $52 \mu \mathrm{~m}$, innermost terminal seta $92 \mu \mathrm{~m}$, and 2 median terminal seta $185 \mu \mathrm{~m}$ (outer) and $220 \mu \mathrm{~m}$ (inner). All setae smooth.

Egg sac containing 1 egg $153 \times 125 \mu \mathrm{~m}$ (Fig. 4e), $135 \times 122 \mu \mathrm{~m}$ (Fig. 4f), $187 \times$ $135 \mu \mathrm{~m}$ (Fig. 4g), 2 eggs $159 \times 140 \mu \mathrm{~m}$ (Fig. 4 h ), 3 eggs (Fig. 4i), or 4 eggs (Fig. 4j).

Rostrum (Fig. 4k) moderately elongate, linguiform. First antenna (Fig. 4l) $280 \mu \mathrm{~m}$ long. Lengths of its 7 segments: 24 ( $55 \mu \mathrm{~m}$ along anterior margin), 88, 23, 26, 40, 27, and $21 \mu \mathrm{~m}$, respectively. Armature: 4, 13, $6,3,4+1$ aesthete, $2+1$ aesthete, and 7 +1 aesthete. All setae smooth.

Second antenna (Fig. 4m) 4-segmented, $185 \mu \mathrm{~m}$ long and moderately robust, second segment $49 \mu \mathrm{~m}$ wide. Armature: 1, 1, 3, and 1 terminal claw $30 \mu \mathrm{~m}$ (Fig. 4 n ).

Labrum (Fig. 5a) triangular with widely separated ventrolateral lobes. Mandible (Fig. 5b) with slender recurved basal part leading to inner lobe with row of slender setules and to outer scalelike area with few small spinules joined to prominent hyaline posteriorly directed process. Lash long and barbed.


Fig. 4. Haplomolgus incolumis, female. a, Dorsal (scale A); b, Urosome, dorsal (D); c, Genital segment, lateral (B); d, Anal segment and caudal ramus, dorsal (E); e, Egg sac, ventral (A); f, Egg sac, ventral (A); g, Egg sac, ventrolateral (A); h, Egg sac, ventral (A); i, Egg sac, ventral (A); j, Egg sac, ventral (A); k, Outline of rostrum, lateral (E); 1, First antenna, with 3 dots showing positions of aesthetes added in male, dorsal (E); m, Second antenna, dorsal (E); n, Claw of second antenna, flat view (F).


Fig. 5. Haplomolgus incolumis, female. a, Labrum and paragnaths, ventral (scale E); b, Mandible, posterior (F); c, First maxilla, outer (F); d, First maxilla, posterior (F); e, Second maxilla, antero-inner (C); f, Maxilliped, postero-outer (C); g, Region between maxillipeds and leg 1, ventral (B); h, Leg 1 and intercoxal plate, anterior (B); i, Outer spine on first segment of exopod of leg 1, anterior (C); j, Leg 2 and intercoxal plate, anterior (B); k, Leg 3 and intercoxal plate, anterior (B); 1, Leg 4 and intercoxal plate, anterior (B).

Paragnath (Fig. 5a) small lobe with several minute spinules. First maxilla (Fig. 5c, d) with 2 smooth setae and 1 barbed spine. Second maxilla (Fig. 5e) with first segment unarmed, second segment with 2 unequal smooth setae and lash with prominent unilateral spines. Maxilliped (Fig. 5f) 3-segmented, first segment unarmed, second segment with 2 small setae, and third segment with 2 unequal setae and 1 terminal spiniform seta.

Ventral area between maxillipeds and first pair of legs not protuberant, with elongate narrow sclerite anterior to intercoxal plate of leg 1 (Fig. 5 g ).

Legs 1-4 (Fig. 5h, j-1) with 3 -segmented rami except for 2 -segmented endopod of leg 4. Formula for armature as follows:

| $P_{1}$ coxa 0-1 basis 1-0 | $\exp \mathrm{I}-0 ; \mathrm{I}-1 ; \mathrm{III}, \mathrm{I}, 4$ |
| :---: | :---: |
|  | enp 0-1; 0-1; I, 2, 3 |
| $\mathrm{P}_{2}$ coxa 0-1 basis 1-0 | $\exp \mathrm{I}-0 ; \mathrm{I}-1 ; \mathrm{III}, \mathrm{I}, 5$ |
|  | enp 0-1; 0-2; I,II, 3 |
| $P_{3}$ coxa 0-1 basis 1-0 | $\exp$ I-0; I-1; II,I,5 |
|  | enp 0-1; 0-2; II, 2 |
| $\mathrm{P}_{4}$ coxa 0-1 basis 1-0 | $\exp \mathrm{I}-0 ; \mathrm{I}-1 ; \mathrm{II}, \mathrm{I}, 5$ |
|  | enp 0-0; I |

Inner margin of coxa in legs $1-3$ with prominent plumose seta but this seta in leg 4 reduced, $18 \mu \mathrm{~m}$ long, and smooth. Exopod of leg 1 with outer spines bearing prominent spinules along proximal edges (Fig. 5i). Exopod of leg $4143 \mu \mathrm{~m}$ long. Endopod (Fig. 6a) with first segment $23.5 \times 18 \mu \mathrm{~m}$ and bearing few outer setules; second segment $21 \times 8 \mu \mathrm{~m}$, with terminal finely barbed spine $39 \mu \mathrm{~m}$.

Leg 5 (Fig. 6b) with free segment elongate oval, $112 \times 47 \mu \mathrm{~m}$, slightly concave inwardly, unornamented, and bearing 2 subterminal smooth setae $55 \mu \mathrm{~m}$ and $61 \mu \mathrm{~m}$. Adjacent smooth dorsal seta $65 \mu \mathrm{~m}$, directed anterodorsally.

Leg 6 represented by 2 minute setae on genital area (Fig. 4c).

Color of living specimens slightly opaque gray, eye red.

Male.-Body (Fig. 6c) resembling that of
female in general form. Length 1.22 mm ( $1.16-1.27 \mathrm{~mm}$ ) and greatest width 0.41 mm ( $0.39-0.44 \mathrm{~mm}$ ), based on 10 specimens. Greatest dorsoventral thickness 0.31 mm . Ratio of length to width of prosome 1.45 : 1. Ratio of length of prosome to that of urosome 1.06:1.

Segment bearing leg 5 (Fig. 6d) $65 \times 164$ $\mu \mathrm{m}$. Genital segment in dorsal view $226 \times$ $244 \mu \mathrm{~m}$, slightly wider than long. Four postgenital segments from anterior to posterior $57 \times 75,62 \times 74,81 \times 65$, and $52 \times 62$ $\mu \mathrm{m}$.

Caudal ramus (Fig. 6d) resembling that of female but smaller, $107 \times 22 \mu \mathrm{~m}$, ratio 4.86:1.

Rostrum like that of female. First antenna segmented and armed as in female, but 3 very long aesthetes added (at points indicated by dots in Fig. 41), so that formula is $4,13+2$ aesthetes, $6,3+1$ aesthete, $4+$ 1 aesthete, $2+1$ aesthete, and $7+1$ aesthete. Longest aesthete, that on proximal part of second segment, $235 \mu \mathrm{~m}$ long (compared with $320 \mu \mathrm{~m}$ for entire length of first antenna). Second antenna (Fig. 6e) resembling that of female, but less robust, width of second segment $39 \mu \mathrm{~m}$.

Labrum, mandible, paragnath, first maxilla, and second maxilla as in female. Maxilliped (Fig. 6f) elongate. First segment unarmed. Second segment with 2 inner setae and group of spines (Fig. 6 g ). Small third segment unarmed. Claw $234 \mu \mathrm{~m}$ long, with 2 unequal proximal setae and bearing minute denticles along slightly concave margin.

Ventral area between maxillipeds and first pair of legs as in female.

Legs $1-4$ similar to those of female, but third segment of endopod of leg 1 with formula I,I,4 (Fig. 6h) with spiniform processes at both sides of inner spine. Leg 4 with endopod like that of female, but in 1 male right endopod with narrow second segment (Fig. 6 i ) and in another male right endopod with formula $0-0 ; 0$ and left endopod with $0-1$; I (Fig. 6j).


Fig. 6. Haplomolgus incolumis. Female: a, Endopod of leg 4, anterior (scale F); b, Leg 5, dorsal (E). Male: c, Dorsal (A); d, Urosome, dorsal (D); e, Second antenna, dorsal (E); f, Maxilliped, postero-inner (B); g, Second segment of maxilliped, inner (E); h, Third segment of endopod of leg 1, anterior (C); i, Pair of abnormal endopods of leg 4, ventral (C); j, Pair of abnormal endopods of leg 4, ventral (C); k, Free segment of leg 5, dorsal (C); 1, Genital and first postgenital segments, ventral (D).

Leg 5 (Fig. 6k) with free segment smaller than in female, $39 \times 21 \mu \mathrm{~m}$, and having few minute spinules along outer border.

Leg 6 (Fig. 61) posteroventral flap on genital segment bearing 2 small setae.

Spermatophore not seen.
Color of living specimens as in female.
Etymology. - The specific name incolum$i s$, Latin meaning unimpaired or undiminished, alludes to the full expression of the endopod of leg 4 in this species.

Remarks. - Two species of Haplomolgus Humes \& Ho, 1968b, have been described, both associated with the scleractinian genus Montipora in Madagascar and Ceram (see Humes \& Ho 1968b, Humes 1978).

Haplomolgus incolumis differs from Haplomolgus montiporae Humes \& Ho, 1968b, in the larger size of the female, the longer female caudal ramus, the endopod of leg 4 with $0-0$; I, and the free segment of the female leg 5 being longer, $112 \times 47 \mu \mathrm{~m}$. Furthermore, in contrast to the new species, $H$. montiporae has 4 setae on the first maxilla and there is only slight sexual dimorphism $(\mathrm{I}, 5)$ in the third segment of the endopod of leg 1. Haplomolgus subdeficiens Humes, 1978, has the formula I, 2 for the third segment of the endopod of leg 3 ; the male of this species is unknown.

The 2 -segmented endopod of leg 4 in Haplomolgus incolumis, armed as $0-0$; I, is similar to the condition in the genus Monomolgus Humes \& Frost, 1964. However, the two genera are distinct in the form of the mandible, the posteriorly directed process in Haplomolgus being absent in Monomolgus.

Haplomolgus montiporae Humes \& Ho, 1968

Specimens studied. - From New Caledonia: 607 \&, 105 oे from Montipora lobulata Bernard, in 0.5 m , Rocher à la Voile, Noumea, $22^{\circ} 18^{\prime} 24^{\prime \prime} \mathrm{S}, 166^{\circ} 25^{\prime} 50^{\prime \prime} \mathrm{E}, 17$ Jun 1971 ; 4 i, 1 oे from Montipora lobulata, in 0.5 m , Rocher à la Voile, Noumea; 136 ㅇ, 129 ô
from Montipora ramosa Bernard, in 2 m , between Isle Noumbo and Isle N'Die, Baie Dumbea, near Noumea, $22^{\circ} 12^{\prime} 47^{\prime \prime}$ S, $166^{\circ} 24^{\prime} 41^{\prime \prime}$ E, 9 Jul 1971; 219 क, 105 o from Montipora ramosa, in 2 m , north of Isle Maître, near Noumea, $22^{\circ} 19^{\prime} 30^{\prime \prime}$ S, $166^{\circ} 24^{\prime} 35^{\prime \prime}$ E, 13 Jul 1971; 62 \&, 24 o from Montipora composita Crossland, in 2 m , Ricaudy Reef, near Noumea, $22^{\circ} 19^{\prime} 00^{\prime \prime}$ S, $166^{\circ} 26^{\prime} 44^{\prime \prime}$ E, 25 Jun 1971.

From northeastern Australia: 16 \&, 5 ô from Montipora composita, in 2 m , Mermaid Cove, Lizard Island, $14^{\circ} 39^{\prime} 50^{\prime \prime} \mathrm{S}$, $145^{\circ} 27^{\prime} 00^{\prime \prime} \mathrm{E}, 27$ Oct 1982; 11 \& , 9 ô from Montipora verrilli Vaughan, in 3 m , Mermaid Cove, Lizard Island, 27 Oct 1982.

Remarks. - Haplomolgus montiporae has been previously reported to be associated with Montipora compressa in the Moluccas (Humes 1978) and with Montipora sinensis, Montipora sp. cf. M. stellata and Montipora sp. in Madagascar (Humes \& Ho 1968b, Humes \& Stock 1973).

Genus Odontomolgus Humes \& Stock, 1972 Odontomolgus forhani Humes, 1978

Specimens studied. - From New Caledonia: 13 \&, 5 ô from Montipora composita Crossland, in 2 m , Ricaudy Reef, near Noumea, $22^{\circ} 19^{\prime} 00^{\prime \prime} \mathrm{S}, 166^{\circ} 26^{\prime} 44^{\prime \prime} \mathrm{E}, 25$ Jun 1971 ; 3 i from Montipora ramosa Bernard, in 2 m , north of Isle Maître, near Noumea, $22^{\circ} 19^{\prime} 30^{\prime \prime} \mathrm{S}, 166^{\circ} 24^{\prime} 35^{\prime \prime} \mathrm{E}, 13 \mathrm{Jul} 1971$; 1 ㅇ from Montipora lobulata Bernard, in 0.5 m , Rocher à la Voile, Noumea, $22^{\circ} 18^{\prime} 24^{\prime \prime} \mathrm{S}$, $166^{\circ} 25^{\prime} 50^{\prime \prime} \mathrm{E}, 17$ Jun 1971.

From the Moluccas: 17 \&, 8 o from Montipora sp. cf. M. undata Bernard, in 10 m , Poelau Parang, eastern Ceram, $03^{\circ} 17^{\prime} 00^{\prime \prime}$ S, 130044ㄴ́"E, 23 May 1975.

From northeastern Australia: 4 is, 1 ô from Montipora composita, in 2 m , Mermaid Cove, Lizard Island, $14^{\circ} 39^{\prime} 50^{\prime \prime} \mathrm{S}$, $145^{\circ} 27^{\prime} 00^{\prime \prime}$ E, 27 Oct 1982; 1 o from Montipora verrilli Vaughan, in 3 m , Mermaid Cove, Lizard Island, 27 Oct 1982.

Remarks.-Odontomolgus forhani has
been previously reported from Montipora compressa and Montipora prolifera in the Moluccas (Humes 1978).

Order Siphonostomatoida Thorell, 1859
Family Asterocheridae Giesbrecht, 1899
Hetairosyna, new genus
Diagnosis. - Asterocheridae. Female: Two postgenital segments. Siphon short, reaching only to maxillipeds. First antenna 19segmented, with aesthete on segment 17. Second antenna with very small 1 -segmented exopod, terminal segment of endopod with claw and 1 seta. Mandible with 1 -segmented palp bearing 1 long seta. Leg 1 with first segment of exopod having inner seta, third segment of exopod having very small proximal vestigial outer spine, with formula III,I,4, or this spine absent, II,I,4. Legs 1-3 with third segment of endopod having I,II, 3, that of segment of leg 4 with I,II,2. Leg 4 with third segment of exopod having III,I,4. Free segment of leg 5 with 3 setae.

Male: Three postgenital segments. First antenna 16 -segmented, with aesthete on segment 15 . Strong sexual dimorphism in endopod of leg 3.

Type species. - Hetairosyna terpna, new species.

Gender. - Feminine.
Etymology. - The generic name Hetairosyna, from the Greek words hetairos, a companion, and the suffix -syne, meaning condition, alludes to the association of members of this genus with corals.

Remarks.-Hetairosyna, though placed in the Asterocheridae following the brief diagnosis given by Stock (1987), differs from the more than 30 genera in the family. Most of these genera may be distinguished from Hetairosyna by means of the key to the genera provided by Stock (1987). Five genera need special attention, however, to distinguish them from the new genus. The salient characters separating them from Hetairosyna are as follows:

Asterocheres Boeck, 1859. The second antenna has three distal elements, the longest more or less setiform; the 1- or 2-segmented palp on the mandible has at least two long setae; and the third segment of the endopod of leg 4 has the formula 1-1 + I-2 (Stock 1987).

Asteropontius Thompson \& A. Scott, 1903. The mandible has a short 1 -segmented palp or the palp is absent, its seta reduced; the third segment of the endopod of leg 4 has 1-1 + I-2 (Stock 1975).

Asteropontella Stock, 1989. The third segment of the exopod of leg 4 has I,I, 3 or II,I, 3; the third segment of the exopod of legs $1-$ 3 has 2 lateral spines; the exopod of the second antenna is vestigial (Stock 1989).

Asteropontoides Stock, 1975. The first antenna of the female has an aesthete on the penultimate segment; in leg 4 the third segment of the endopod has $1, \mathrm{I}, 2$ and the third segment of the exopod has III,I, 3 (Sewell 1949, Stock 1975).

Asteropontopsis Stock, 1987. The third segment of the exopod of leg 3 has II,I,4; the third segment of the endopod of leg 3 has $1,1+\mathrm{I}, 3$ (Stock 1987).

Hetairosyna terpna new species
Figs. 7, 8, 9, 10
Type material. - 43 ㅇ, 40 i from Montipora caliculata (Dana), in 1.5 m , Ricaudy Reef, near Noumea, New Caledonia, $22^{\circ} 19^{\prime} 00^{\prime \prime} \mathrm{S}, 166^{\circ} 26^{\prime} 44^{\prime \prime} \mathrm{E}, 25$ Jun 1971. Holotype $\&$ (USNM 254462), allotype (USNM 254463), and 75 paratypes ( 38 \& 37 ) (USNM 254461) deposited in the National Museum of Natural History, Smithsonian Institution, Washington, D.C. Remaining paratypes (dissected) in the collection of the author.

Other specimens. - From New Caledonia: 11 \& from Montipora composita Crossland, in 2 m , Ricaudy Reef, near Noumea, 25 Jun 1971; 3 \& from Montipora ramosa Bernard, in 2 m , north of Isle Maître, near Noumea, $22^{\circ} 19^{\prime} 30^{\prime \prime}$ S, $166^{\circ} 24^{\prime} 35^{\prime \prime}$ E, 13 Jul 1971; 30
\&, 13 î from Montipora lobulata Bernard, in 0.5 m , Rocher à la Voile, Noumea, $22^{\circ} 18^{\prime} 24^{\prime \prime}$ S, $166^{\circ} 25^{\prime} 50^{\prime \prime}$ E, 17 Jun 1971; 93 $\ddagger$ from Montipora sp. cf. M. efflorescens Bernard, in 1 m , east of Isle N'Die, Baie Dumbea, near Noumea, $22^{\circ} 13^{\prime} 15^{\prime \prime}$ S, $166^{\circ} 24^{\prime} 26^{\prime \prime}$ E, 6 Jul 1971.

From the Moluccas: 14 from Montipora compressa (Esper), in 2 m , Natsepa, Ambon, $03^{\circ} 37^{\prime} 05^{\prime \prime} \mathrm{S}, 128^{\circ} 17^{\prime} 00^{\prime \prime} \mathrm{E}, 28$ May 1975 ; 3 \& from Montipora prolifera Brueggemann, in 3 m , southern shore of Goenoeng Api, Banda Islands, $04^{\circ} 32^{\prime} 05^{\prime \prime} \mathrm{S}, 129^{\circ} 52^{\prime} 30^{\prime \prime} \mathrm{E}, 26$ Apr 1975.

From Enewetak Atoll, Marshall Islands: 17 \& from Montipora ambigua Bernard, in 3 m , south of Parry Island, 9 Jul 1969; 12 $\%$ from Montipora patula Bernard, in 2 m , western end of Bogen Island, 23 Jun 1969.

From northeastern Australia: 8 \&, 6 o from Montipora composita, in 2 m , Mermaid Cove, Lizard Island, $14^{\circ} 39^{\prime} 50^{\prime \prime} \mathrm{S}$, $145^{\circ} 27^{\prime} 00^{\prime \prime}$ E, 27 Oct 1982.

From Madagascar: 41 \&, 4 i from Montipora sp., west of Pte de Tafondro, Nosy Bé, 3 Dec 1963; 3 \& from Montipora sinensis Bernard, in 1 m , Nosy Taolankena, near Nosy Bé, 15 Nov 1963; 12 i, 2 ô from Montipora sp. cf. M. stellata Bernard, in 2 m , off Ampombilava, Nosy Bé, 26 Sept 1964.

Female. - Body (Fig. 7a) with broad prosome covered with minute hyaline scales. Length $0.92 \mathrm{~mm}(0.88-1.01 \mathrm{~mm})$ and greatest width $0.49 \mathrm{~mm}(0.47-0.52 \mathrm{~mm})$, based on 10 specimens. Greatest dorsoventral thickness 0.33 mm . Epimera of segments bearing legs $1-4$ rounded in dorsal view, weakly pointed in lateral view. Dorsal part of segment bearing leg 3 partly covering segment bearing leg 4 . Segment bearing leg 1 not separated dorsally from cephalosome. Ratio of length to width of prosome 1.29: 1. Ratio of length of prosome to that of urosome 2.09:1.

Segment bearing leg 5 (Fig. 7b) $60 \times 152$ $\mu \mathrm{m}$. Genital segment slightly expanded in anterior third, $153 \times 127 \mu \mathrm{~m}$ in greatest dimensions in dorsal view, $91 \mu \mathrm{~m}$ wide in
posterior third. Genital areas located dorsolaterally in anterior third. Each area (Fig. 7b-d) with 2 very small setae. Surfaces of all urosomal segments covered with minute hyaline scales (Fig. 7b-d). Genital segment with row of long setules laterally posterior to expansion (Fig. 7d). Two postgenital segments, first segment $62 \times 78 \mu \mathrm{~m}$, second segment $50 \times 75 \mu \mathrm{~m}$.

Caudal ramus (Fig. 7e) short, trapezoidal, wider than long, $29 \mu \mathrm{~m}$ along outer edge, $18 \mu \mathrm{~m}$ along inner edge, and $36 \mu \mathrm{~m}$ in greatest width. Outer lateral seta placed dorsally, $107 \mu \mathrm{~m}$, dorsal seta $72 \mu \mathrm{~m}$, both smooth. Outermost terminal seta $156 \mu \mathrm{~m}$, innermost terminal seta $195 \mu \mathrm{~m}$, and 2 median terminal setae $273 \mu \mathrm{~m}$ (outer) and $330 \mu \mathrm{~m}$ (inner), swollen proximally. Ramus bearing few outer scales as on urosomal segments.

Dorsal surface of body without visible sensilla, but with minute scales.

Egg sac (Fig. 7f) $252 \times 190 \mu \mathrm{~m}$, containing 5 eggs, each about $135-140 \mu \mathrm{~m}$ in maximum diameter.

Rostral area weakly developed (Fig. 8a). First antenna (Fig. 8b) $418 \mu \mathrm{~m}$ long, 19segmented. Length of segments: 23 ( $47 \mu \mathrm{~m}$ along anterior margin), 13, 10, 10, 10, 10, $11,13,10,15.5,26,25,29,27,29,34,37$, 14 , and $26 \mu \mathrm{~m}$, respectively. Formula for armature: $1,2,2,2,2,2,2,2,6,1+1$ small spine, $2,2,2,2,2,2,2+1$ aesthete, 3, and 7. All setae smooth.

Second antenna (Fig. 8c) 5-segmented, $257 \mu \mathrm{~m}$ long including claw. Coxa and basis unarmed. Exopod minute, $7 \times 4 \mu \mathrm{~m}$. Endopod with first segment elongate and unarmed, second segment with 1 seta, third segment with terminal claw $57 \mu \mathrm{~m}, 1$ adjacent seta, and outer row of setules.

Siphon (Fig. 8d) short, approximately 165 $\mu \mathrm{m}$ long. Mandible (Fig. 8e) with long slender blade $143 \mu \mathrm{~m}$, its tip dentate inwardly and expanded outwardly (Fig. 8f); palp 1 -segmented with long slender seta, palp and seta together $91 \mu \mathrm{~m}$. Both mandibles lying along sides of siphon (Fig. 8d). First maxilla (Fig. 8g) with 2 lobes, shorter outer lobe


Fig. 7. Hetairosyna terpna, female. a, Dorsal (scale A); b, Urosome, dorsal (B); c, Urosome, lateral (B); d, Genital segment, dorsal (E); e, Anal segment and caudal ramus, dorsal (C); f, Egg sac, ventral (D). (In a-e scales not shown over entire surface.)


Fig. 8. Hetairosyna terpna, female. a, Rostral area, ventral (scale B); b, First antenna, anterodorsal (E); c, Second antenna (E); d, Siphon and mandibles, ventral (B); e, Mandible, anterior (E); f, Tip of mandible, posteroinner (F); g, First maxilla, posterior (E); h, Second maxilla, inner (E); i, Maxilliped, posterior (E); j, Leg 1 and intercoxal plate, anterior (B).
with 4 setae, longer inner lobe with 4 setae, 1 distinctly barbed. Second maxilla (Fig. 8h) 2 -segmented, first segment unarmed, second segment clawlike with small seta at midlength. Maxilliped (Fig. 8i) 5-segmented, first and second segments unarmed, third, four, and fifth segments with 1 seta each, claw $70 \mu \mathrm{~m}$.

Ventral region between maxillipeds and first pair of legs not protuberant.

Legs $1-4$ (Figs. 8j, 9a-c) biramous, with 3 -segmented rami. Formula for armature as follows:

$$
\begin{aligned}
& P_{1} \text { coxa 0-1 basis 1-I } \exp \text { I-1; I-1; III,I,4 } \\
& \text { enp 0-1; 0-2; 1,2,3 } \\
& \mathrm{P}_{2} \text { coxa 0-1 basis 1-0 } \exp \text { I-1; I-1; III,I,4 } \\
& \text { enp 0-1; 0-2; 1,II,3 } \\
& \exp \text { I-1; I-1; III,I,4 } \\
& \text { enp 0-1; 0-2; 1,II,3 } \\
& \mathrm{P}_{4} \text { coxa 0-1 basis 1-0 } \exp \mathrm{I}-1 ; \mathrm{I}-1 ; \text { III,I,4 } \\
& \text { enp 0-1; 0-2; 1,II, } 2
\end{aligned}
$$

Legs 1-3 with long plumose seta on inner side of coxa, but this seta reduced in leg 4. Inner element on basis setiform spine. Inner margin of basis in leg 1 with minute marginal spinules, but smooth in legs $2-4$. Proximal outer spine on third segment of exopod of leg 1 very small, vestigial. In 1 female, right endopod of leg 1 with aberrant formula $0-2 ; 0-2 ; 1,2,3$.

Leg 5 (Fig. 9d) with free segment $36 \times$ $15.5 \mu \mathrm{~m}$, with few minute scales on outer margin, its 3 setae 20,55 , and $65 \mu \mathrm{~m}$, respectively. Adjacent dorsal seta $52 \mu \mathrm{~m}$. All setae smooth.

Leg 6 represented by 2 minute setae on genital area (Fig. 7d).

Color of living specimens in transmitted light transparent to opaque gray, eye red.

Male. - Body (Fig. 9e) with prosome more slender than in female. Length 0.73 mm ( $0.70-0.78 \mathrm{~mm}$ ) and greatest width 0.33 mm ( $0.31-0.35 \mathrm{~mm}$ ), based on 10 specimens. Greatest dorsoventral thickness 0.25 mm . Epimeral areas slightly more pointed than in female. Ratio of length to width of prosome 1.49:1. Ratio of length of prosome to
that of urosome 1.86:1. Urosomal segments with surficial scales as in female.

Caudal ramus (Fig. 9f) resembling that of female but smaller, $18 \mu \mathrm{~m}$ along outer edge, $11 \mu \mathrm{~m}$ along inner edge, and $26 \mu \mathrm{~m}$ wide.

Body surface as in female.
Rostral area like that of female. First antenna (Fig. 9g) $340 \mu \mathrm{~m}$ long, 16 -segmented (though segment 10 possibly divided).
Lengths of segments: 23 ( $38 \mu \mathrm{~m}$ along anterior margin), $13,12,12,12,12,12,12$, $12,16,48,15.5,13,40,39$, and $34 \mu \mathrm{~m}$, respectively. Armature: 1, 2, 2, 2, 2, 2, 2 , $2,6,1+1$ small spine, $2,1,1,0,0+1$ aesthete, and 7. Setae hyaline and difficult to observe.

Second antenna, siphon, mandible, first maxilla, and second maxilla as in female. Maxilliped (Fig. 9h) resembling that of female but showing sexual dimorphism in having inner spiniform process on both first and second segments.

Ventral area between maxillipeds and first pair of legs as in female.

Legs $1-4$ segmented and armed as in female, but sexual dimorphism in legs 2 and 3. Endopod of leg 2 (Fig. 10a) with terminal spiniform process on third segment larger than in female. Endopod of leg 3 (Fig. 10b) with second segment having enlarged terminal outer process; third segment distinctly smaller than first two, with formula 1,II,3, but having enlarged swollen inner terminal process bearing 1 outer setule.

Leg 5 (Fig. 10c) with minute free segment $13 \times 10 \mu \mathrm{~m}$, bearing 3 setae as in female.

Leg 6 (Fig. 10d) posteroventral flap on genital segment bearing 2 setae $10 \mu \mathrm{~m}$ and $26 \mu \mathrm{~m}$.

Spermatophore unknown.
Color as in female.
Etymology. - The specific name terpna is derived from the Greek word terpnos, meaning delightful, pleasing.

Remarks.-Hetairosyna terpna is associated with at least 11 species of Montipora from Madagascar to Enewetak Atoll to New Caledonia (Table 3).


Fig. 9. Hetairosyna terpna. Female: a, Leg 2 and intercoxal plate, anterior (scale B); b, Leg 3 and intercoxal plate, anterior (B); c, Leg 4 and intercoxal plate, anterior (B); d, Leg 5, dorsal (C). Male: e, Dorsal (A); f, Urosome, lateral (B); g, First antenna, anterodorsal (E); h, Maxilliped, posterior (E).


Fig. 10. Hetairosyna terpna, male. a, Endopod of leg 2, anterior (scale C); b, Endopod of leg 3, anterior (C); c, Leg 5, ventral (F); d, Genital segment, ventral (B).

Hetairosyna laciniata, new species Figs. 11, 12, 13

Type material. - 11 \&, 6 ơ from Montipora caliculata (Dana), in 1.5 m , Ricaudy Reef, near Noumea, New Caledonia, $22^{\circ} 19^{\prime} 00^{\prime \prime} \mathrm{S}$, $166^{\circ} 26^{\prime} 44^{\prime \prime} \mathrm{E}, 25$ Jun 1971. Holotype ㅇ (USNM 254467), allotype (USNM 254468), and 11 paratypes ( 7 \&, 4 ठ) (USNM 254466) deposited in the National Museum of Natural History, Smithsonian Institution, Washington, D.C. Remaining paratypes (dissected) in the collection of the author.

Other specimens. -From New Caledonia: 16 \&, 5 ô from Montipora composita, in 2 m, Ricaudy Reef, near Noumea, 25 Jun 1971; 19 ㅇ, 17 ô from Montipora ramosa Bernard, in 2 m , north of Isle Maître, near Noumea, $22^{\circ} 19^{\prime} 30^{\prime \prime} \mathrm{S}, 166^{\circ} 24^{\prime} 35^{\prime \prime} \mathrm{E}$, 13 Jul 1971; 27 i, 41 ô from Montipora lobulata Bernard, in 0.5 m , Rocher à la Voile, Noumea, $22^{\circ} 18^{\prime} 24^{\prime \prime} \mathrm{S}, 166^{\circ} 25^{\prime} 50^{\prime \prime} \mathrm{E}, 17$ Jun 1971.

From Madagascar. - 2 \&, 2 o from Montipora sinensis Bernard, in 1 m , Nosy Taolankena, near Nosy Bé, 15 Nov 1963.

Female. - Body (Fig. 11a) with unusually broad orbicular prosome. Length 0.87 mm $(0.81-0.90 \mathrm{~mm})$ and greatest width 0.58 mm ( $0.56-0.61 \mathrm{~mm}$ ), based on 10 specimens.

Greatest dorsoventral thickness 0.24 mm . Segment bearing leg 1 fused with cephalosome and finely serrate on lateral margins. Epimera of segments bearing legs $2-4$ truncate and finely and irregularly serrate. Dorsal surface of prosome tessellated (Fig. 11b). Ratio of length to width of prosome 1.03: 1. Ratio of length of prosome to that of urosome 1.86:1.

Segment bearing leg 5 (Fig. 11c) $73 \times 150$ $\mu \mathrm{m}$. Genital segment $134 \times 138 \mu \mathrm{~m}$, nearly as long as wide in dorsal view, widest in anterior half. Row of several slender setules on both sides near middle of segment. Dorsal surface of segment with mostly crescentic sculpturing (Fig. 11d). Genital areas located dorsolaterally at expanded portion of segment. Each area with 2 minute setae and very small spinous process (Fig. 11d). Two postgenital segments, first segment $61 \times 65$ $\mu \mathrm{m}$, second segment $55 \times 57 \mu \mathrm{~m}$, both with delicate minute scales laterally and continuing over dorsal surfaces.

Caudal ramus (Fig. 11e) short, $22 \mu \mathrm{~m}$ long on outer side, $19 \mu \mathrm{~m}$ long on inner side, and $24 \mu \mathrm{~m}$ wide. Outer lateral seta (placed dorsally) $20 \mu \mathrm{~m}$, dorsal seta $20 \mu \mathrm{~m}$, outermost terminal seta $44 \mu \mathrm{~m}$, innermost terminal


Fig. 11. Hetairosyna laciniata, female. a, Dorsal (scale A); b, Tessellations on surface of prosome, dorsal (E); c, Urosome, ventral (B); d, Genital segment, dorsal (B); e, Anal segment and caudal ramus, dorsal (C); f, First antenna, posteroventral (E); g, Second antenna, antero-outer (E); h, Rostral area and siphon, ventral (A); i, Mandible, anterior (B); j, Tip of mandible, flat view (C).
setae $73 \mu \mathrm{~m}$, and 2 long median terminal setae, outer $570 \mu \mathrm{~m}$, and inner (swollen proximally) $930 \mu \mathrm{~m}$. All setae smooth. Few scales on outer surface of ramus.

Dorsal surface of body without visible sensilla, but prosome tessellated (Fig. 11b).

Egg sac not seen.
Rostral area weak (Fig. 11h). First antenna (Fig. 11f) $340 \mu \mathrm{~m}$ long, 19 -segmented. Lengths of segments 26 ( $52 \mu \mathrm{~m}$ along anterior margin), $15,11,10,10,11,10,13$, $13,13,14,14,18,21,22,22,28,14$, and $19 \mu \mathrm{~m}$, respectively. Armature: 1, 2, 2, 2, $2,2,2,2,6,1+1$ small spine, $2,2,2,2$, $2,2,2+1$ aesthete, 3, and 7. All setae smooth.

Second antenna (Fig. 11 g ) probably 5 -segmented, though third segment of endopod obscure. Coxa and basis unarmed. Exopod minute, $2.5 \times 2.5 \mu \mathrm{~m}$, with 1 seta. Endopod with first segment elongate with small knob on proximal outer corner and with few distal outer setules. Second segment triangular. Third segment with terminal claw $56 \mu \mathrm{~m}$, 1 adjacent seta, and outer marginal setules.

Siphon (Fig. 11h) short, reaching only to bases of maxillipeds. Mandible (Fig. 11i) with long slender blade $160 \mu \mathrm{~m}$, its tip dentate and expanded inwardly (Fig. 11j). Palp 1 -segmented with long smooth seta, palp and seta together $164 \mu \mathrm{~m}$. First maxilla (Fig. 12a) with both lobes bearing 4 setae, 2 setae on longer inner lobe barbed. Second maxilla (Fig. 12b) 2-segmented, first segment unarmed, second slender recurved segment clawlike, with faint indication of division midway, at that point bearing 1 seta and few very small spinules. Maxilliped (Fig. 12c) 5 -segmented, first and second segments unarmed, third, fourth, and fifth segments with 1 seta each, claw $60 \mu \mathrm{~m}$, with abruptly slender tip preceded by 1 minute inner spinule.

Ventral region between maxillipeds and first pair of legs not protuberant.

Legs $1-4$ (Fig. 12d, e, g, i) biramous, with 3 -segmented rami. Formula for armature as follows:

| $\mathrm{P}_{1}$ coxa 0-1 basis 1-I | $\exp$ I-0; I-1; III,I,4 |
| :---: | :---: |
|  | enp 0-1; 0-2; 1,2,3 |
| $\mathrm{P}_{2}$ coxa 0-1 basis 1-0 | $\exp$ I-1; I-1; III,I,4 |
|  | enp 0-1; 0-2; 1,II,3 |
| $\mathrm{P}_{3}$ coxa 0-1 basis 1-0 | $\exp$ I-1; I-1; III,I,4 |
|  | enp 0-1; 0-2; 1,II,3 |
| $\mathrm{P}_{4}$ coxa 0-1 basis 1-0 | $\exp$ I-1; I-1; III,I,4 |
|  | enp 0-1; 0-2; 1,II, 2 |

Inner coxal seta of leg 1 short, $13 \mu \mathrm{~m}$, and smooth, that of leg 2 long and weakly plumose, that of leg 3 short as in leg 1 , and that of leg 4 very short, $5 \mu \mathrm{~m}$. Inner spine on basis of leg $144 \mu \mathrm{~m}$, coarsely barbed along outer side and with setules along inner side (Fig. 12d). Legs 2-4 with few small spines on distal outer corner of coxa and on outer margin of basis. Inner margin of basis in leg 1 with row of small spines, in legs $2-4$ this margin smooth. First segment of exopod of leg 1 with outer spine long and recurved, inner seta absent; in legs 2-4 this spine shorter and not recurved. Leg 2 in 1 female with endopod having formula $0-1 ; 0-2 ; 1, \mathrm{II}, 2$ (Fig. 12f). Leg 3 in 1 female with endopod having 0-1; 0-2; 1,II, 2 (Fig. 12h).

Leg 5 (Fig. 13a) with small subrectangular unornamented free segment $18 \times 10.5 \mu \mathrm{~m}$, its 3 setae from outer to inner 27,25 , and $8 \mu \mathrm{~m}$. Adjacent dorsal seta $30 \mu \mathrm{~m}$.

Leg 6 represented by 2 minute setae on genital area (Fig. 11d).

Color of living specimens in transmitted light opaque gray, eye red.

Male. - Body (Fig. 13b) with broad prosome slightly less orbicular than in female. Length $0.71 \mathrm{~mm}(0.65-0.75 \mathrm{~mm})$ and greatest width $0.43 \mathrm{~mm}(0.42-0.45 \mathrm{~mm})$, based on 6 specimens. Greatest dorsoventral thickness 0.18 mm . Epimera of segments bearing legs $1-4$ serrate as in female. Ratio of length to width of prosome 1.12:1. Ratio of length of prosome to that of urosome 1.90:1.

Segment bearing leg 5 (Fig. 13c) $21 \times 116$ $\mu \mathrm{m}$. Genital segment $104 \times 150 \mu \mathrm{~m}$, much wider than long. Length including leg 6109 $\mu \mathrm{m}$. Segment rounded laterally and bearing small scales. Three postgenital segments


Fig. 12. Hetairosyna laciniata, female. a, First maxilla, posterior (scale E); b, Second maxilla, posterior (B); c, Maxilliped, posterior (B); d, Leg 1 and intercoxal plate, anterior (E); e, Leg 2 and intercoxal plate, anterior (E); f, Endopod of leg 2, anterior (E); g, Leg 3 and intercoxal plate, anterior (E); h, Endopod of leg 3, anterior (E); i, Leg 4 and intercoxal plate, anterior (E).


Leg 3 with endopod showing sexual dimorphism (Fig. 13f), with spiniform process on second segment much longer than in female, third segment with distal outer process enlarged and sinuous and outer seta somewhat spiniform.

Leg 5 (Fig. 13 g ) with very small free segment $10 \times 6 \mu \mathrm{~m}$.

Leg 6 (Fig. 13h) posteroventral flap on genital segment bearing 2 small setae 26 and $17 \mu \mathrm{~m}$.

Spermatophore (Fig. 13i) oval, $63 \times 48$ $\mu \mathrm{m}$.

Color of living specimens as in female.
Etymology. - The specific name lacinia$t a$, Latin meaning fringed, refers to the finely serrated fringe on the epimera of the prosomal segments.

Remarks. - Hetairosyna laciniata may be recognized by the serrated fringe on the epimera of the prosomal segments in both sexes. In addition, the female of the new species shows several features that distinguish it from H. terpna: (1) the prosome is broader and more orbicular, (2) the female genital segment is as wide as long, (3) the seta on the palp of the mandible is smooth, (4) the second segment of the maxilliped is relatively shorter, (5) the inner coxal seta in legs $1-4$ is short, and (6) the free segment of leg 5 is twice as long. In the male of $H$. laciniata the genital segment is much wider than in H. terpna.

Hetairosyna sororia, new species Figs. 14, 15, 16

Type material. - 10 \& from Montipora ramosa Bernard, in 2 m , north of Isle Maître, near Noumea, New Caledonia, $22^{\circ} 19^{\prime} 30^{\prime \prime} \mathrm{S}$, $166^{\circ} 24^{\prime} 35^{\prime \prime}$ E, 13 Jul 1971. Holotype (USNM 254436) and 7 paratypes (USNM 254437) deposited in the National Museum of Natural History, Smithsonian Institution, Washington, D.C. Remaining paratypes (dissected) in the collection of the author.

Other specimens. - From Madagascar, 1 $\circ$ from Montipora sinensis Bernard, in 1 m ,

Nosy Taolankena, near Nosy Bé, 15 Nov 1963; 1 \& from Montipora sp. cf. M. stellata Bernard, in 2 m , off Ampombilava, Nosy Bé, 26 Sep 1964.

From northeastern Australia: 2 ifrom Montipora verrilli Vaughan, in 3 m , Mermaid Cove, Lizard Island, $14^{\circ} 39^{\prime} 50^{\prime \prime} \mathrm{S}$, $145^{\circ} 27^{\prime} 00^{\prime \prime} \mathrm{E}, 27$ Oct 1982.

From the Moluccas: 2 ifrom Montipora sp. cf. M. undata Bernard, Poelau Parang, near Ceram, $03^{\circ} 17^{\prime} 00^{\prime \prime}$ S, $130^{\circ} 44^{\prime} 48^{\prime \prime} \mathrm{E}, 23$ May 1975.

Female. - Body (Fig. 14a) with broad prosome. Length $0.99 \mathrm{~mm}(0.94-1.01 \mathrm{~mm})$ and greatest width 0.55 mm ( $0.50-0.61$ mm ), based on 10 specimens. Greatest dorsoventral thickness 0.30 mm . Epimera of segments bearing legs $1-3$ pointed posteriorly, those of segment bearing leg 4 subtruncate. Ratio of length to width of prosome 1.26:1. Ratio of length of prosome to that of urosome 2.35:1.

Segment bearing leg 5 (Fig. 14b) $57 \times 177$ $\mu \mathrm{m}$. Genital segment quadrate, $148 \times 151$ $\mu \mathrm{m}$ in dorsal view, with slightly convex sides. Genital areas situated dorsolaterally near middle of segment. Each genital area with 2 minute setae and spiniform process (Fig. $14 \mathrm{~b}, \mathrm{c}$ ). Row of long strong setules on outer margin of segment posterior to genital areas. Two postgenital segments, first segment 62 $\times 92 \mu \mathrm{~m}$, second segment $62 \times 87 \mu \mathrm{~m}$.

Caudal ramus (Fig. 14d) $29 \times 36 \mu \mathrm{~m}$ in greatest dimensions, wider than long. Outer lateral seta, placed dorsally as in congeners, $107 \mu \mathrm{~m}$, dorsal seta $65 \mu \mathrm{~m}$, both smooth. Outermost terminal seta $165 \mu \mathrm{~m}$, innermost terminal seta $195 \mu \mathrm{~m}$, and 2 median terminal setae $330 \mu \mathrm{~m}$ (outer) and $380 \mu \mathrm{~m}$ (inner), swollen proximally, all with lateral setules.

Dorsal surface of prosome with extremely small scales. Urosome with more prominent scales (Fig. 14b) extending over dorsal and ventral surfaces (Fig. 14b,c) (all scales not illustrated). Caudal rami with few inner and outer scales (Fig. 14d).

Egg sac not seen.


Fig. 14. Hetairosyna sororia, female. a, Dorsal (scale A); b, Urosome, dorsal (B); c, Segment bearing leg 5 and genital segment, lateral (B); d, Anal segment and caudal ramus, dorsal (E); e, First antenna, ventral (B). (In a-d scales not shown over entire surface.)

Rostral area weak, slightly protruding ventrally (Fig. 15b). First antenna (Fig. 14e) $460 \mu \mathrm{~m}$ long, 19 -segmented. Lengths of segments: 26 ( $62 \mu \mathrm{~m}$ along anterior margin), $15.5,13,11,11,11,11,14,15,14,31,29$, $31,29,31,36,43,15$, and $30 \mu \mathrm{~m}$, respectively. Formula for armature: $1,2,2,2,2$, $2,2,2,6,1+1$ small spine, $2,2,2,2,2$, $2,2+1$ aesthete, 3 , and 7 . All setae smooth. First segment with small spinules along convex margin.

Second antenna (Fig. 15a) 5-segmented, $352 \mu \mathrm{~m}$ long including claw. Coxa and basis unarmed. Exopod minute, $10 \times 5 \mu \mathrm{~m}$. Endopod with first segment elongate and unarmed, second segment with 1 small seta, and third segment with terminal claw 115 $\mu \mathrm{m}, 1$ adjacent seta, and outer marginal row of setules.

Siphon (Fig. 15b, c) moderately slender, $180 \mu \mathrm{~m}$ long, reaching posteriorly beyond bases of maxillipeds. Mandible (Fig. 15d) with long slender blade $211 \mu \mathrm{~m}$, its tip expanded outwardly as in Hetairosyna terpna; palp 1-segmented with long smooth terminal seta, palp and seta together approximately $200 \mu \mathrm{~m}$ long. First maxilla (Fig. 15e) with 2 unequal lobes, both with 4 setae, 2 of these setae on larger lobe with lateral setules; larger lobe with row of indistinct setules on anterior surface. Second maxilla (Fig. 15f) 2-segmented, first segment unarmed, second segment with minute seta proximal to middle of segment. Maxilliped (Fig. 15g) 5 -segmented, armed as in H. terpna, claw $104 \mu \mathrm{~m}$.

Ventral region between maxillipeds and first pair of legs protuberant (Fig. 15c).

Legs 1-4 (Figs. 15h, i, 16a, b) segmented as in congeners. Formula for armature as follows:

| $\mathrm{P}_{1}$ coxa 0-1 basis 1-I | $\exp \mathrm{I}-1 ; \mathrm{I}-1 ; \mathrm{II}, \mathrm{I}, 4$ |
| :---: | :---: |
|  | enp 0-1; 0-2; 1,5 |
| $\mathrm{P}_{2}$ coxa 0-1 basis 1-0 | $\exp$ I-1; I-1; III,I,4 |
|  | $\text { enp } 0-1 ; 0-2 ; 1, \mathrm{II}, 3$ |
| $\mathrm{P}_{3}$ coxa 0-1 basis 1-0 | $\begin{array}{ll} \exp \text { I-1; I-1; III,I,4 } \\ \text { enp 0-1; } 0-2 ; 1, I I, 3 \end{array}$ |
| $\mathrm{P}_{4}$ coxa 0-1 basis 1-0 | $\exp$ I-1; I-1; III,I,4 |
|  | enp 0-1; 0-2; 1,II,2 |

Inner spine on basis of leg $152 \mu \mathrm{~m}$ in length with long setules (Fig. 15h). Leg 1 with third segment of exopod having II,I,4, vestigial spine seen in congeners here absent. Inner coxal seta on leg 4 short, $16 \mu \mathrm{~m}$.

Leg 5 (Fig. 16c) with free segment $47 \times$ $23 \mu \mathrm{~m}$, ornamented with few small outer spinules, bearing from outer to inner 3 setae 65,55 , and $18 \mu \mathrm{~m}$. Adjacent dorsal seta 78 $\mu \mathrm{m}$. All setae smooth.

Leg 6 represented by 2 minute setae on genital area (Fig. 14c). Color of living specimens in transmitted light transparent to opaque gray, eye red.

Male. - Unknown.
Etymology. - The specific name sororia, Latin meaning sisterly, alludes to the occurrence of this species along with two other species of Hetairosyna.

Remarks.-Hetairosyna sororia may be recognized by five features: (1) the siphon reaching posteriorly beyond the bases of the maxillipeds, (2) the strong setules on the lateral margins of the genital segment, (3) the lateral setules on the terminal setae of the caudal ramus, (4) the small spinules on the convex margin of the first segment of the first antenna, (5) the third segment of the exopod of leg 1 with II,I,4.

The three species of Hetairosyna may be differentiated as shown in Table 1.

## Tychomyzon, new genus

Diagnosis. - Asterocheridae. Female: Two postgenital segments. Siphon relatively short. First antenna 18 -segmented, with aesthete on segment 17 . Second antenna with very small exopod, terminal segment of endopod with 1 claw and 1 seta. Mandible with 1 -segmented palp bearing long seta. Leg 1 with first segment of endopod having inner seta. Leg 1 with inner coxal seta but this seta absent in legs 2-4. Third segment of endopod in legs $1-3$ with formula $1,2,3$, but in leg 4 with $1,2,1$. Leg 4 with third segment of exopod having II,I,4. Leg 5 with minute free segment bearing 3 setae.

Male: Three postgenital segments. First antenna 16-segmented with aesthete on seg-


Fig. 15. Hetairosyna sororia, female. a, Second antenna, antero-outer (scale B); b, Cephalosome, ventral (A); c, Part of cephalosome, lateral (D); d, Mandible, anterior (B); e, First maxilla, posterior (E); f, Second maxilla, inner (B); g, Maxilliped, posterior (B); h, Leg 1 and intercoxal plate, anterior (B); i, Leg 2 and intercoxal plate, anterior (B).


Fig. 16. Hetairosyna sororia, female. a, Leg 3 and intercoxal plate, anterior (scale B); b, Leg 4 and intercoxal plate, anterior (B); c, Leg 5, dorsal (C).
ment 15. Sexual dimorphism not expressed in legs 1-4.

Type species. - Tychomyzon petalum, new species.

Gender. - Neuter.
Etymology. - The generic name Tychomyzon, a combination of the Greek words tyche, good fortune, and myzon, used in the
names of several asterocherid genera and derived from myzao, to suck.

Tychomyzon petalum, new species
Figs. 17, 18, 19
Type material. - 7 \&, 1 oे from Montipora composita Crossland, in 3 m , Mermaid Cove, Lizard Island, northeastern Austra-

Table 1.-Differentiation of the three species of Hetairosyna, based on females.

|  | H. terpna | H. laciniata | H. sororia |
| :--- | :--- | :--- | :--- |
| Prosome, ratio L/W | $1.29: 1$ | $1.03: 1$ | $1.26: 1$ |
| Prosomal epimera | smooth weakly | fringed pointed | smooth pointed |
|  | rounded |  |  |
| Caudal ramus, terminal setae | with setules | smooth | with setules |
| Genital segment, dorsal view | $153 \times 127 \mu \mathrm{~m}$ | $134 \times 138 \mu \mathrm{~m}$ | $148 \times 151 \mu \mathrm{~m}$ |
| Genital segment, setules on sides | weak hairlike | weak hairlike | strong |
| Genital segment, dorsal surface | smooth | crescentic | smooth |
|  |  | sculpturing |  |
| First antenna, anterior margin of | smooth | smooth | with spinules |
| first segment |  |  |  |
| Siphon | reaches bases of | reaches bases of | reaches posterior to |
|  | maxillipeds | maxillipeds | bases of maxillipeds |
| Leg 1 exopod, third segment | III,I,4 | III,I,4 | II,I,4 |
| Leg 5, free segment | $36 \times 15.5 \mu \mathrm{~m}$ | $18 \times 10.5 \mu \mathrm{~m}$ | $47 \times 23 \mu \mathrm{~m}$ |



Fig. 17. Tychomyzon petalum, female. a, Dorsal (scale A); b, Lateral (A); c, Urosome, ventral (B); d, Genital segment, dorsal (D); e, Genital segment, ventral (D); f, Genital segment, lateral (D); g, Anal segment and caudal ramus, dorsal (C); h, Tesselations on dorsal surface of cephalosome, dorsal (E); i, Detail of tessellation on dorsal surface of cephalosome, dorsal (F); j, Egg sac, ventral (D); k, Rostral area and siphon, ventral (B); 1, First antenna, posteroventral (C).


Fig. 18. Tychomyzon petalum, female: a, Second antenna, antero-outer (scale C); b, Siphon, dorsal (E); c, Mandible, anterior (E); d, First maxilla, anterior (E); e, Second maxilla, anterior (E); f, Maxilliped, posterior (E); g , Leg 1 and intercoxal plate, anterior (E); h, Leg 2 and intercoxal plate, anterior (E); i, Leg 3 and intercoxal plate, anterior (E).


Fig. 19. Tychomyzon petalum. Female: a, Leg 4 and intercoxal plate, posterior (scale A); b, Leg 5, ventral (F). Male: c, Dorsal (A); d, Urosome, ventral (E); e, First antenna, anterodorsal (C).
lia, $14^{\circ} 39^{\prime} 50^{\prime \prime} \mathrm{S}, 145^{\circ} 27^{\prime} 00^{\prime \prime} \mathrm{E}, 27$ Oct 1982. Holotype $\&$ (USNM 254442), allotype (USNM 254441), and 4 paratype 9 (USNM 254473) deposited in the National Museum of Natural History, Smithsonian Institution, Washington, D.C. Remaining paratypes (dissected) in the collection of the author.

Female. - Body (Fig. 17a, b) with broad prosome, segment bearing leg 4 hidden in dorsal view beneath more anterior segment (Fig. 17b). Length $0.74 \mathrm{~mm}(0.68-0.78 \mathrm{~mm})$ and greatest width 0.51 mm (0.49-0.52 mm ), based on 7 specimens. Greatest dorsoventral thickness 0.26 mm . Epimera of segment bearing leg 1 pointed, those of segment bearing leg 2 rounded and finely serrated. In dorsal view segment bearing leg 3 triangular and segment bearing leg 4 small,
without expanded epimera. Ratio of length to width of prosome 1.19:1. Ratio of length of prosome to that of urosome 1.94:1.

Segment bearing leg 5 (Fig. 17c) $39 \times 100$ $\mu \mathrm{m}$. Genital segment $109 \times 88 \mu \mathrm{~m}$, longer than wide, in dorsal view broadest in anterior half. Genital areas situated dorsolaterally, both bearing 2 small setae (Fig. 17d). Ventral surface with median opening of sperm duct (Fig. 17e, f).

Caudal ramus (Fig. 17g) small, subquadrate, $18 \mu \mathrm{~m}$ in outer length, $15 \mu \mathrm{~m}$ in inner length, and $18 \mu \mathrm{~m}$ wide. Outer lateral seta $20 \mu \mathrm{~m}$ (displaced dorsally), dorsal seta 22 $\mu \mathrm{m}$, outermost terminal seta $38 \mu \mathrm{~m}$, innermost terminal seta $30 \mu \mathrm{~m}$, and 2 long median terminal setae $133 \mu \mathrm{~m}$ (outer) and 195 $\mu \mathrm{m}$ (inner). All setae smooth.

Dorsal surface of prosome with irregu-

Table 2.-Montipora in the Indo-Pacific and associated Copepoda. E = Enewetak Atoll, Marshall Islands, $\mathrm{H}=$ Hawaii, $\mathrm{NC}=$ New Caledonia, $\mathrm{A}=$ northeastern Australia, MOL=Moluccas, $\mathrm{I}=\mathrm{India}$, MAD = Madagascar, MAU = Mauritius.

| Montipora ambigua Bernard |  |
| :---: | :---: |
| Hetairosyna terpna, new species | E |
| Montipora caliculata (Dana) |  |
| Euryte bellatula, new species | NC |
| Haplomolgus incolumis, new species | NC |
| Haplomolgus terpna, new species | NC |
| Montipora composita Crossland |  |
| Haplomolgus montiporae Humes \& Ho, 1968b | NC |
| Hetairosyna laciniata, new genus, new species | NC |
| Hetairosyna terpna, new species | A, NC |
| Odontomolgus forhani Humes, 1978 (present paper) | A, NC |
| Tychomyzon petalum, new genus, new species | A |
| Xarifia anopla Humes \& Dojiri, 1982 (in Humes 1985) | A |
| Xarifia heteromeles Humes \& Dojiri, 1982 (in Humes 1985) | A |
| Montipora compressa (Esper) |  |
| Euryte bellatula, new species | MOL |
| Haplomolgus montiporae Humes \& Ho, 1968b (in Humes 1978) | MOL |
| Hetairosyna terpna, new species | MOL |
| Odontomolgus forhani Humes, 1978 | MOL |
| Montipora sp. cf. M. efflorescens Bernard |  |
| Hetairosyna terpna, new species | NC |
| Montipora foliosa (Pallas) |  |
| Indoclausia bacescui Sebastian \& Pillai, 1974 (host reported as M. foliacea, but see |  |
| Montipora lobulata Bernard |  |
| Euryte bellatula, new species | NC |
| Haplomolgus montiporae Humes \& Ho, 1968b (present paper) | NC |
| Hetairosyna laciniata, new species | NC |
| Hetairosyna terpna, new species | NC |
| Odontomolgus forhani Humes, 1978 (present paper) | NC |
| Montipora patula Verrill |  |
| Hetairosyna terpna, new species | E |
| Montipora prolifera Brueggemann |  |
| Euryte bellatula, new species | MOL |
| Hetairosyna terpna, new species | MOL |
| Odontomolgus forhani Humes, 1978 | MOL |
| Montipora ramosa Bernard |  |
| Euryte bellatula, new species | NC |
| Haplomolgus montiporae Humes \& Ho, 1968b (present paper) | NC |
| Hetairosyna laciniata, new species | NC |
| Hetairosyna sororia, new species | NC |
| Hetairosyna terpna, new species | NC |
| Odontomolgus forhani Humes, 1978 (present paper) | NC |
| Xarifia pectinea Humes \& Dojiri, 1982 | NC |
| Xarifia temnura Humes \& Ho, 1968a (in Humes \& Dojiri 1982) | NC |

Table 2.-Continued.

| Montipora sinensis Bernard |  |
| :---: | :---: |
| Haplomolgus montiporae Humes \& Ho, 1968b | MAD |
| Hetairosyna laciniata, new species | MAD |
| Hetairosyna sororia, new species | MAD |
| Hetairosyna terpna, new species | MAD |
| Xarifia temnura Humes \& Ho, 1968a | MAD |
| Montipora sp. cf. M. stellata Bernard |  |
| Haplomolgus montiporae Humes \& Ho, 1968b | MAD |
| Hetairosyna sororia, new species | MAD |
| Hetairosyna terpna, new species | MAD |
| Xarifia sp. (in Humes 1985) | MAD |
| Montipora sp. cf. M. undata Bernard |  |
| Allopodion mirum Humes, 1978 | MOL |
| Haplomolgus subdeficiens Humes, 1978 | MOL |
| Hetairosyna sororia, new species | MOL |
| Kawanolus parangensis Humes, 1978 | MOL |
| Odontomolgus forhani Humes, 1978 (present paper) | MOL |
| Xarifia anopla Humes \& Dojiri, 1982 | MOL |
| Xarifia heteromeles Humes \& Dojiri, 1982 | MOL |
| Xarifia syntoma Humes \& Dojiri, 1982 | MOL |
| Xarifia temnura Humes \& Ho, 1968a (in Humes \& Dojiri 1982) | MOL |
| Montipora verrilli Vaughan |  |
| Alteuthellopsis corallina Humes, 1981 (in Humes 1984) | A |
| Euryte bellatula, new species | A |
| Haplomolgus montiporae Humes \& Ho, 1968b (present paper) | A |
| Hetairosyna sororia, new species | A |
| Odontomolgus forhani Humes, 1978 (present paper) | A |
| Montipora verrucosa (Lamarck) |  |
| Tegastes gemmeus Humes, 1984 | H |
| Xarifia apertipes Humes \& Dojiri, 1983 | MAD |
| Montipora sp. |  |
| Asteropontius corallophilus Stock, 1966 | MAU |
| Haplomolgus montiporae Humes \& Ho, 1968b (also in Humes \& Stock 1973) | MAD |
| Hetairosyna terpna, new species | MAD |
| Xarifia anopla Humes \& Dojiri, 1982 | MAD |
| Xarifia extensa Humes \& Dojiri, 1982 | A, MAD |

larly shaped tessellations having minute slender digitiform processes (Fig. 17h, i).

Egg sac (Fig. 17j) with 2 eggs, both approximately $130 \times 117 \mu \mathrm{~m}$.

Rostrum (Fig. 17k) small and inconspicuous. First antenna (Fig. 171) $240 \mu \mathrm{~m}$ long, 18 -segmented. Lengths of segments: 16 (32 $\mu \mathrm{m}$ along anterior margin), $11,8,8,5,5,8$, $10,12,10,12,12,12,12,12,16.5,20$, and
$23 \mu \mathrm{~m}$, respectively. Armature: 1, 2, 2, 2 , $2,2,2,2,6,1+1$ small spine, $1,1,1,1$, $1,2,2+1$ aesthete, and 6 . All setae smooth and hyaline.

Second antenna (Fig. 18a) 5-segmented, $195 \mu \mathrm{~m}$ long. Coxa and basis unarmed. Exopod minute knob, $2 \mu \mathrm{~m}$ long, without visible setae. Endopod with elongate first segment bearing inner setules, short triangular

Table 3.-Copepoda associated with Montipora in the Indo-Pacific.

| Allopodion mirum | Montipora compressa |
| :---: | :---: |
| Montipora sp. cf. M. undata | Montipora sp. cf. M. efflorescens |
| Alteuthellopsis corallina | Montipora lobulata |
| Montipora verrilli | Montipora patula |
| Asteropontius corallophilus | Montipora prolifera |
| Montipora sp. | Montipora ramosa |
| Euryte bellatula, new species | Montipora sinensis |
| Montipora caliculata | Montipora sp. cf. M. stellata |
| Montipora compressa | Montipora sp. |
| Montipora lobulata | Odontomolgus forhani |
| Montipora prolifera | Montipora composita |
| Montipora ramosa | Montipora lobulata |
| Montipora verrilli | Montipora ramosa |
| Haplomolgus incolumis, new species | Montipora sp. cf. M. undata |
| Montipora caliculata | Montipora verrilli |
| Haplomolgus montiporae | Tegastes gemmeus |
| Montipora composita | Montipora verrucosa |
| Montipora compressa | Tychomyzon petalum, new genus, new species |
| Montipora lobulata | Montipora composita |
| Montipora ramosa | Xarifia anopla |
| Montipora sp. cf. M. stellata | Montipora composita |
| Montipora sinensis | Montipora sp. cf. M. undata |
| Montipora verrilli | Montipora sp. |
| Montipora sp. | Xarifia apertipes |
| Haplomolgus subdeficiens | Montipora verrucosa |
| Montipora sp. cf. M. undata | Xarifia extensa |
| Hetairosyna laciniata, new genus, new species | Montipora sp. |
| Montipora composita | Xarifia heteromeles |
| Montipora lobulata | Montipora composita |
| Montipora ramosa | Montipora sp. cf. M. undata |
| Montipora sinensis | Xarifia pectinea |
| Hetairosyna sororia, new genus, new species | Montipora ramosa |
| Montipora ramosa | Xarifia syntoma |
| Montipora sinensis | Montipora sp. cf. M. undata |
| Montipora sp. cf. M. undata | Xarifia temnura |
| Montipora verrilli | Montipora ramosa |
| Hetairosyna terpna, new genus, new species | Montipora sinensis |
| Montipora ambigua | Montipora sp. cf. M. undata |
| Montipora caliculata | Xarifia sp. |
| Montipora composita | Montipora sp. cf. M. stellata |

second segment with 1 seta, and third segment bearing terminally 1 claw $34 \mu \mathrm{~m}$ long and 1 seta.

Siphon (Figs. 17k, 18b) $150 \mu \mathrm{~m}$ long. Mandible (Fig. 18c) with long slender blade $109 \mu \mathrm{~m}$, its tip inwardly dentate and expanded outwardly. Palp 1 -segmented with long smooth seta, palp and seta together 117 $\mu \mathrm{m}$ long. First maxilla (Fig. 18d) with 2 unequal lobes, longer inner lobe with inner
margin having setules and bearing 4 terminal setae, one of them unilaterally with setules; shorter outer lobe with 4 setae, one of them subterminal. Second maxilla (Fig. 18e) 2-segmented, first segment unarmed, second segment recurved with 1 minute seta midway on posterior surface. Maxilliped (Fig. 18f) 5-segmented, first 2 segments unarmed, last 3 segments with 1 seta each, terminal claw $47 \mu \mathrm{~m}$ with beaklike tip.

Legs 1-4 (Figs. 18g-i, 19a) biramous and 3 -segmented. Formula for armature as follows:

```
P
    enp 0-1; 0-1; 1,2,3
P
    enp 0-1; 0-2; 1,2,3
P
    enp 0-1; 0-2; 1,2,3
P
    enp 0-1; 0-2; 1,2,1
```

Leg 1 with inner coxal seta, but this seta absent in legs 2-4. Basis of leg 1 having inner barbed spine $26 \mu \mathrm{~m}$ long, inner expansion of basis with row of small denticles on margin. Outer spine on first segment of exopod of leg 1 enlarged, recurved, $31 \mu \mathrm{~m}$ long. Leg 4 with third segment of endopod having $1,2,1$.

Leg 5 (Figs. 17c, 19b) minute, free segment $13 \times 9 \mu \mathrm{~m}$, unornamented and inserted slightly ventrally. Free segment armed with 3 setae.

Leg 6 represented by 2 small setae on genital segment (Fig. 17d).
Color of living specimens in transmitted light opaque gray, eye red.
Male. - Body (Fig. 19c) with broad prosome. Length 0.51 mm , greatest width 0.40 mm , based on allotype. Segment bearing leg 4 not covered in dorsal view by segment bearing leg 3. Ratio of length to width of prosome 1:1. Ratio of length of prosome to that of urosome 1.53:1.
Segment bearing leg 5 (Fig. 19d) $31 \times 90$ $\mu \mathrm{m}$. Genital segment $80 \times 120 \mu \mathrm{~m}$, wider than long, widest in anterior third. Three postgenital segments from anterior to posterior $28 \times 46,37 \times 39$, and $39 \times 37 \mu \mathrm{~m}$.

Caudal ramus like that of female but smaller, $13 \times 13 \mu \mathrm{~m}$.
Dorsal surface of prosome not tessellated as in female.

Rostrum similar to that of female. First antenna (Fig. 19e) $195 \mu \mathrm{~m}$ long, 16 -segmented, slightly modified. Lengths of segments: 13 ( $33 \mu \mathrm{~m}$ along anterior margin),
$10,9,4,5,6,8,9,10,10,19,9,9,21,24$, and $14 \mu \mathrm{~m}$, respectively. Armature: $1,2,2$, $2,2,2,2,2,6,1+1$ small spine, $1,1,1$, $1,1+1$ aesthete, and 6 . Setae smooth and hyaline.

Second antenna, siphon, mandible, first maxilla, second maxilla, maxilliped, and legs 1-4 resembling those of female.

Leg 5 (Fig. 19d) with minute free segment approximately $8 \times 6 \mu \mathrm{~m}$.

Leg 6 (Fig. 19d) posteroventral flap on genital segment bearing 2 small setae.

Extruded spermatophore not seen.
Color as in female.
Etymology. - The specific name petalum, Greek petalos, broad, outspread, alludes to the unusually broad prosome.

Relatively few species of Montipora have been examined for copepod associates. The 15 species of Montipora recorded as hosts in Table 2 constitute less than half of the 38 species known from Australia listed by Veron (1986), and only a small fraction of the 211 nominal species. Ten species of Montipora have three or more species of copepod associates, with the greatest number being nine in the case of Montipora sp. cf. M. undata in the Moluccas. With 20 species of copepods already known to live with Montipora (Table 3), it seems certain that further studies will reveal many more copepods associated with this large and abundant coral genus.

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## Literature Cited

Boeck, A. 1859. Tvende nye parasitiske Krebsdyr, Artotrogus orbicularis og Asterocheres liljebor-gii.-Forhandlinger i Videnskabs-Selskabet i Christiania 1859:171-182.
Burmeister, H. 1834. Beiträge zur Naturgeschichte der Rankenfüsser (Cirripedia). G. Reimer, Berlin. Pp. 1-59.
Dana, J. D. 1853. Crustacea. In United States Exploring Expedition during the years 1838-1842 under the command of Charles Wilkes 13(1):iviii, 1-685, 13(2):686-1618.
Giesbrecht, W. 1899. Die Asterocheriden des Golfes von Neapel. - Fauna und Flora des Golfes von Neapel 25:i-vi, 1-217.
-_. 1900. Mittheilungen über Copepoden. 12-14.- Mittheilungen aus der Zoologischen Station zu Neapel 14:39-82.
Humes, A. G. 1978. Lichomolgid copepods (Cyclopoda) associated with the coral genus Montipora in the Moluccas. - Publications of the Seto Marine Biological Laboratory 24:376-407.
1981. Harpacticoid copepods associated with Cnidaria in the Indo-West Pacific. - Journal of Crustacean Biology 1:227-240.
1984. Harpacticoid copepods associated with cnidarians in the tropical Pacific Ocean.-Zoologica Scripta 13:209-221.
1985. A review of the Xarifidae (Copepoda, Poecilostomatoida), parasites of scleractinian corals in the Indo-Pacific.-Bulletin of Marine Science 36:467-632.
, \& M. Dojiri. 1982. Xarifidae (Copepoda) parasitic in Indo-Pacific scleractinian corals.Beaufortia 32:139-228.
, \& ——. 1983. Copepoda (Xarifidae) parasitic in scleractinian corals from the Indo-Pa-cific.-Journal of Natural History 17:257-307. , \& B. W. Frost. 1964. New lichomolgid co-
pepods (Cyclopoida) associated with alcyonarians and madreporarians in Madagascar.-Cahiers ORSTOM Océanographie, 1963, 6(série Nosy Bé II):131-212.
, \& R. U. Gooding. 1964. A method for studying the external anatomy of copepods. - Crustaceana 6:238-240.
, and J.-S. Ho. 1968a. Xarifid copepods (Cyclopoida) parasitic in corals in Madagascar. Bulletin of the Museum of Comparative Zoology 136:415-460.
, \& ——. 1968b. Lichmolgid copepods (Cyclopoida) associated with corals in Madagas-car.-Bulletin of the Museum of Comparative Zoology 136:353-413.
, \& J. H. Stock. 1972. Preliminary notes on a revision of the Lichomolgidae, cyclopoid copepods mainly associated with marine inver-tebrates.-Bulletin Zoologisch Museum, Universiteit van Amsterdam 2:121-133.
, \& - . 1973. A revision of the Lichomolgidae, cyclopoid copepods mainly associated with marine invertebrates.-Smithsonian Contributions to Zoology 127:i-v, 1-368.
Kossmann, R. 1877. Entomostraca (1. Theil: Lichomolgidae). Pp. 1-24 in Zoologische Ergebnisse einer im Auftrage der Königlichen Academie der Wissenschaften zu Berlin aufgeführten Reise in die Küstensgebiete des Rothen Meeres, erste Hälfte, IV.
Philippi, A. 1843. Fernere Beobachtungen über die Copepoden des Mittlemeeres. - Archiv für Na turgeschichte 9:54-71.
Sars, G. O. 1913. An account of the Crustacea of Norway with short descriptions and figures of all the species. Pp. 1-32. in Vol. VI Copepoda Cyclopoida Parts I \& II Oithonidae, Cyclopinidae, Cyclopinae (part). Bergen Museum, Alb. Cammermeyer's Forlag, Christiania, Norway.
Scott, T. 1912. The Entomostraca of the Scottish National Antarctic Expedition, 1902-1904.Transactions of the Royal Society of Edinburgh 48:521-599.
Sebastian, M. J., \& N. K. Pillai. 1974. Two new genera of clausiid copepods, Indoclausia and Stockia.--Crustaceana 26:80-88.
Sewell, R. B. S. 1949. The littoral and semi-parasitic Cyclopoida, the Monstrilloida and the Notodelphyoida. John Murray Expedition 1933-34, Scientific Reports, 13(2):17-199.
Stock, J. H. 1966. Cyclopoida siphonostoma from Mauritius (Crustacea, Copepoda).-Beaufortia 13:145-194.
1975. Copepoda associated with West Indian Actiniaria and Corallimorpharia.-Studies on the Fauna of Curaçao and other Caribbean Islands 48:88-118.
-_. 1987. Copepoda Siphonostomatoida asso-
ciated with West Indian hermatypic corals 1: associates of Scleractinia: Faviinae. - Bulletin of Marine Science 40:464-483.
1989. Copepoda Siphonostomatoidea associated with West Indian hermatypic corals. 2.Studies in honour of Dr. Pieter Wagenaar Hummelinck, Foundation for Scientific Research in Surinam and the Netherlands Antilles, Amsterdam 123:145-169.
Thompson, I. C., \& A. Scott. 1903. Report on the Copepoda collected by Professor Herdman, at Ceylon, in 1902.-Report to the Government of Ceylon on the Pearl Oyster Fisheries of the Gulf of Manaar, supplementary reports 7:227307.

Thorell, T. 1859. Till kannedomen om vissa parasitiskt lefvande Entomostraceer.-Ofversift af Kongl. Vetenskaps-Akademiens Forhandlingar 16:333-362.

Ummerkutty, A. N. P. 1970. The genus Euryte Philippi (Copepoda, Cyclopoida) in Indian waters. - Records of the Zoological Survey of India, 1966 64:113-119.
Veron, J. E. N. 1986. Pp. i-xii, 1-664 in Corals of Australia and the Indo-Pacific. Angus \& Robertson Publishers, North Ryde, New South Wales, Australia.
Vervoort, W. 1964. Free-living Copepoda from Ifaluk Atoll in the Caroline Islands with notes on related species.-United States National Museum Bulletin 236:i-ix, 1-431.

Boston University Marine Program, Marine Biological Laboratory, Woods Hole, Massachusetts 02543.


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