

Choreftria shiranui, a new genus and species of cyclopoid copepod (Crustacea: Copepoda) associated with the worm goby from southern Japan, with the proposal of a new family

Daisuke Uyeno

Received: 19 August 2021/Accepted: 3 November 2021/Published online: 24 January 2022 © The Author(s), under exclusive licence to Springer Nature B.V. 2022

Abstract A new genus and species of cyclopoid copepod, Choreftria shiranui n. g., n. sp. (Crustacea: Copepoda) is described based on an adult female found from the worm goby, Taenioides snyderi Jordan & Hubbs (Actinopterygii: Perciformes: Gobiidae), from mud flats in the Yatsushiro Sea, southern Japan. The new genus is characterized by bearing the following characters in the female: body distinctly segmented cyclopiform with 4-segmented prosome and 6-segmented urosome; genital somite clearly separated from first abdominal somite; antennule 7-segmented; antenna 4-segmented with one fused serrated claw and long claw on second endopodal segment; mandible reduced with one serrated blade; maxillule rod-like with one element; maxilla 2-segmented with recurved terminal claw; maxilliped 2-segmented; legs 1 to 4 biramous with 3-segmented rami. Since the copepod is not attributable to any of the known cyclopoid families, a new family, Choreftriidae **n. fam.** is established to accommodate *Choreftria* **g.** LSID urn:lsid:zoobank.org:pub:C1FB9F60-9871-4D4B-A1BF-3202BA24189F.

Graduate School of Science and Engineering, Kagoshima University, 1-21-35 Korimoto, Kagoshima 890-0065, Japan

e-mail: duyeno@sci.kagoshima-u.ac.jp

Introduction

Marine fishes are the major hosts of parasitic copepods (Crustacea: Copepoda). Of the order Cyclopoida, members of the following seven families are known to inhabit marine fish: Bomolochidae Claus, 1875, Chondracanthidae Milne Edwards, 1840, Makrostrotidae Huys et al., 2012, Philichthyidae Vogt, 1877, Shiinoidae Cressey, 1975, Taeniacanthidae Wilson C.B., 1911, and Telsidae Ho, 1967 (Boxshall & Halsey, 2004; Huys et al., 2012). Several members of the two families Ergasilidae Burmeister, 1835 and Lernaeidae Cobbold, 1879, of which members are commonly found from fresh- to brackish waters, also occur on marine fishes. Additionally, one species from each of the following families, Anthessiidae Humes, 1986, Pseudanthessiidae Humes & Stock, 1972, and Macrochironidae Humes & Boxshall, 1996, was found from marine fishes, while other members of those families were found on invertebrate hosts (Avdeev, 1975; Avdeev & Kazachenko, 1986; Boxshall & Halsey, 2004; Uyeno & Rain, 2021).

In this study, a new genus and species of cyclopoid copepod that is not attributable to the known families is described based on an adult female attached to the worm goby *Taenioides snyderi* Jordan & Hubbs (Actinopterygii: Perciformes: Gobiidae) collected from the Yatsushiro Sea, southern Japan. A new family is established to accommodate this new genus.



D. Uyeno (⊠)

Materials and methods

A fish was collected from at low tide on the mud flats in the Yatsushiro Sea using a yabby pump. A specimen of copepod was removed from the host's anal fin in the laboratory, fixed and preserved in 80% ethanol. Subsequently, the copepod specimen was soaked in lactophenol for 24 h, dissected with a sharpened insect pin under a dissecting microscope (Leica M80), and observed, based on a modified procedure of the wooden slide method of Humes & Gooding (1964), under a biological microscope (Olympus BX53). Illustrations were made with the aid of a drawing tube attached to the biological microscope. The copepod's body was measured using an ocular micrometer; measurements are given in µm. The type specimen of the copepod and its host fish are deposited in the crustacean collection of the Natural Museum of Nature of Science, Tsukuba, Japan (NSMT-Cr) and The Kagoshima University Museum, Kagoshima City, Japan (KAUM-I), respectively. The scientific and common names of fishes used in this paper follow FishBase (Froese & Pauly, 2021).

Results

Choreftria n. g.

LSID urn:lsid:zoobank.org:act:37FF9F26-96F1-444B-B344-F88FE691FDB5

New Japanese name: Oshiri-kajiri-mushi-zoku.

Diagnosis

Adult female. Body cyclopiform. Second to fifth pedigerous somites, genital somite and four abdominal somites free. Genital somite with genital opening on ventral surface. Abdomen composed of four free somites. Caudal ramus with one outer, one inner, and four apical setae.

Rostrum absent. Antennule 7-segmented; aesthetascs present on sixth and distal segments. Antenna 4-segmented, composed of coxobasis and 3-segmented endopod; coxobasis large, bearing single naked distal seta; proximal endopodal segment bearing naked seta on inner median margin; second endopodal segment bearing one serrated fused distal claw, one long apical claw, and one pectinate and one

naked setae; distal endopodal segment slightly laterally displaced, bearing one subterminal and six terminal setae. Labrum with serrated median linguiform process on posterior margin. Mandible composed of robust basal part and elongate curved distal part bearing blade with three posterior denticles. Paragnath ovoid with smooth margin. Maxillule represented by small rod with one apical blunt element. Maxilla 2-segmented; proximal segment unarmed; distal claw recurved. Maxilliped 2-segmented; proximal segment rod-shaped without armature; distal segment rod-shaped bearing small apical element.

Legs 1 to 4 biramous, each with 3-segmented rami. Intercoxal sclerite present on legs 1 to 4. Leg 5 2-segmented; proximal segment bearing dorsal seta and row of fine spinules on distal margin; distal segment bearing one outer and two distal naked setae and rows of fine spinules near bases of setae. Leg 6 represented by three long setae at genital opening.

Adult male. Unknown.

Type-species: Choreftria shiranui **n. sp.**, by original designation.

Etymology: The new generic name is derived from the Greek noun, χορεθτρια, which means "dancer", making reference to the well-developed long leg 4 of the new genus which is reminiscent of a dancer's leg. The gender is feminine.

Remarks

Choreftria n. g. has the following unusual characters on the cephalothorax: 1) the second endopodal segment of the antenna is armed with one fused claw and one long claw, and distal endopodal segment is laterally displaced; 2) the elongate mandible is reduced with a serrated apical blade; and 3) the maxilla is robust with a curved terminal claw. Of cyclopoid copepods, three genera, Leptinogaster Pelseneer, 1929, *Pholadicola* Ho & Wardle, 1992, and Teredicola C. B. Wilson, 1942 which are defined as members of the Teredicola-group by Boxshall & Halsey (2004) share the similar form of antenna and maxilla (see Wilson, 1957; Bocquet & Stock, 1958; Humes & Turner, 1972; Ho & Wardle, 1992; Kim, 2009). Especially, *Leptinogaster* is similar to the new genus in sharing an untransformed body with four well



developed pairs of swimming legs composed of coxae, bases, and both rami 3-segmented (e.g. Humes & Cressey, 1958; Kim & Ho, 1991; Kim, 2009). However, *Choreftria* n. g. is distinguished by the possession of a reduced mandible with only the terminal blade while those three other genera each bear two more elements in addition to the apical blade. *Gadilicola* Boxshall & O'Reilly, 2015, the type genus of the monotypic family Gadilicolidae Boxshall & O'Reilly, 2015 shares the similar antenna, mandible, and maxilla with the new genus (see Boxshall & O'Reilly, 2015). However, the body and swimming legs of *Gadilicola* are highly transformed while those of the new genus are not transformed, without distinct reduction.

Choreftria shiranui n. g., n. sp.

LSID urn:lsid:zoobank.org:act:589A498B-96B5-4043-BADF-9D6ACD189D95

New Japanese name: Oshiri-kajiri-mushi.

Type-host: Taenioides snyderi Jordan & Hubbs (Actinopterygii: Perciformes: Gobiidae) (KAUM–I–156767, 158.6 mm SL).

Type-locality: Off the mouth of the Kojiro River (32°06'23.9"N, 130°18'23.4"E), Izumi, Kagoshima, the Yatsushiro Sea, Japan.

Attachment site: Surface of the fin membrane of the anal fin (Fig. 1).

Type-material: Holotype, adult female (NSMT-Cr 30590), collected by Reo Koreeda from tidal flat at a depth of 0 m, 1 May 2021.



Fig. 1 Fresh coloration of holotype adult female of *Choreftria shiranui* **n. g., n. sp.** (NSMT-Cr 30590) attached on the anal fin of *Taenioides snyderi* Jordan & Hubbs (KAUM–I–156767, 158.6 mm SL), photographed by Reo Koreeda

Etymology: The specific name "shiranui" refers to the name of both the natural phenomenon and folklore creature around the type locality, the Yatsushiro Sea. Further, the alias of the type locality is also the Shiranui Sea.

Description (Figs. 2-4)

Adult female

Body (Fig. 2A) cyclopiform, 1370 long, with greatest width at cephalothorax and second pedigerous somite. Cephalosome and first pedigerous somite forming cephalothorax. Cephalothorax wider than long, 334×436 . Second to fifth pedigerous somites, genital somite, and four abdominal somites free. Prosome 664 long. Genital somite oval, wider than long, 137×248 , slightly constricted at anterior third; genital opening situated on ventral surface (Fig. 2B). Abdomen composed of four free somites, 86×96 , 75×98 , 69×95 , and 112×85 , respectively. Caudal ramus (Fig. 2A, C) 4.84 times longer than wide, 188×39 , with vertical dorsal ridge near posterior end and one outer, one inner, and four apical setae.

Rostrum absent. Antennule (Fig. 2D) 7-segmented; armature formula 4, 13, 4, 3, 4, 2 + 1 aesthetasc, 7 + 1aesthetasc; all setae naked, except one plumose seta on posterior margin. Antenna (Fig. 2E) 4-segmented, composed of coxobasis and 3-segmented endopod; coxobasis large, bearing single naked distal seta; proximal endopodal segment bearing naked seta on inner median margin; second endopodal segment bearing one serrated fused distal claw, one long apical claw, and one pectinate and one naked setae; distal endopodal segment slightly laterally displaced bearing one subterminal and six terminal setae. Labrum (Fig. 3A, B) with serrated median linguiform process on posterior margin. Mandible (Fig. 3A, C, D) composed of robust basal part and elongate curved distal part bearing blade with three posterior denticles (Fig. 3D). Paragnath (Fig. 3A, E) ovoid with smooth margin. Maxillule (Fig. 3A, F) represented by small rod with one apical blunt element. Maxilla (Fig. 3A, G) 2-segmented; proximal segment unarmed; distal claw recurved. Maxilliped (Fig. 3A, H) 2-segmented; proximal segment rod-shaped without armature; distal segment rod-shaped bearing small apical element.

Legs 1 to 4 (Figs. 4A–D) biramous, each with 3-segmented rami. Armature formula as follows



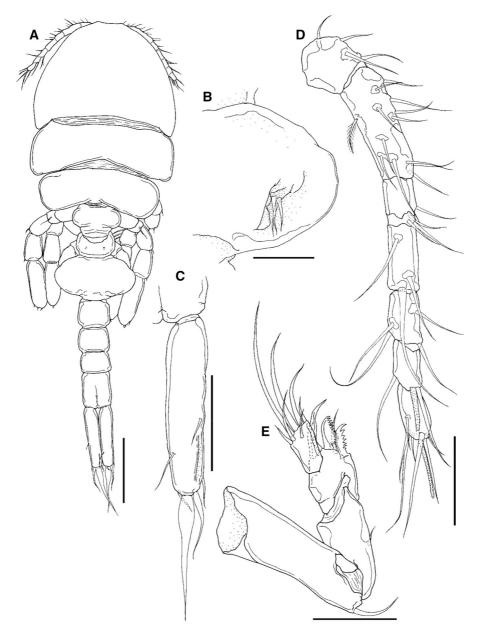


Fig. 2 *Choreftria shiranui* **n. g., n. sp.**, adult female, holotype, NSMT-Cr 30590. A, habitus, dorsal; B, partial genital somite, left side, ventral; C, right caudal ramus, dorsal; D, left antennule, posterior; E. right antenna, posterior. Scale bars: A, 200 μm; B, D, 50 μm; C, 100 μm; E, 30 μm

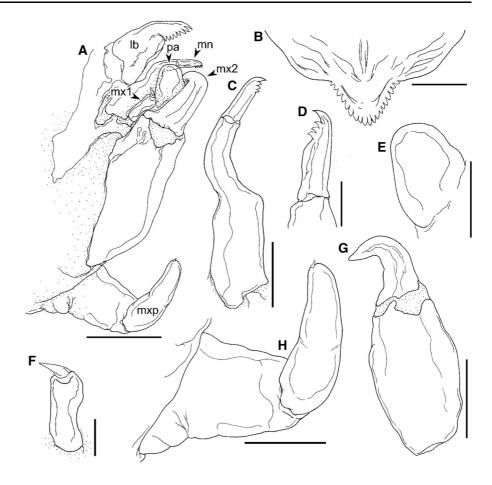
(Arabic numbers = number of seta, Roman numbers = number of spines):

	Coxa	Basis	Exopod	Endopod
Leg 1	0-1	1-I	I-0; I-1; III, 1, 4	0-1; 0-1; I, 2, 3
Leg 2	0-1	1-0	I-0; I-1; III, 2, 4	0-1; 0-2; II, I, 3

	Coxa	Basis	Exopod	Endopod
Leg 3	0-1	1-0	I-0; I-1; II, 1, 4	0-1; 0-2; I, 2
Leg 4	0-0	1-0	1-0; 1-1; 1, 2	0-1; 0-1; 1



Fig. 3 Choreftria shiranui n. g., n. sp., adult female, holotype, NSMT-Cr 30590. A, oral region, right side, lateral, lb = labrum, mn = mandible, pa = paragnath, mx1 = maxillule, mx2 =maxilla, mxp = maxilliped;B, labrum, anterior; C, right mandible, outer; D, distal tip of right mandible, inner; E, right paragnath; F, right maxillule, anterior; G, left maxilla, posterior; H, right maxilliped, outer. Scale bars: A, G, 50 µm; B, C, E, 20 μm; D, F, 10 μm; H, 30 μm



Intercoxal sclerite of leg 1 (Fig. 4A) with pair of marginal rows of hairs. Intercoxal sclerites of legs 2 to 4 (Figs. 4B–D) with pair of posterior rows of spinules. Coxae, bases, and rami of legs 1 to 4 bearing row of spinules on posterior margins. Middle endopodal segments of legs 1 to 3 each armed with pointed distal projection on outer margin. All setae on legs 1 to 3, except on bases, plumose. Leg 5 (Fig. 4E) 2-segmented; proximal segment bearing dorsal seta and row of fine spinules on distal margin; distal segment bearing one outer and two distal naked setae and rows of fine spinules near bases of setae. Leg 6 (Fig. 2B) represented by three long setae at genital opening.

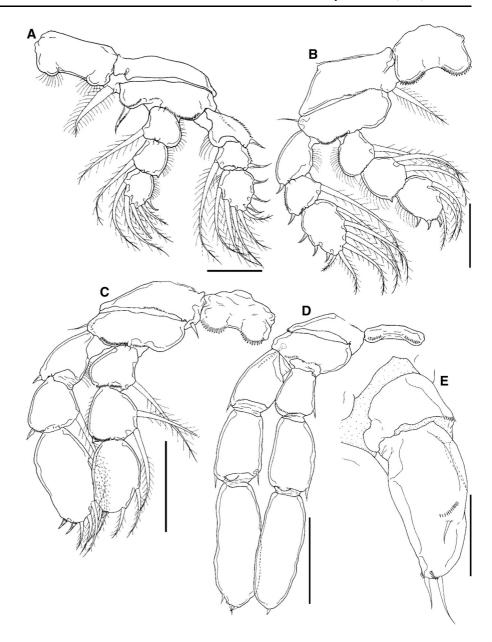
Discussion

As commented in the Remarks section of this paper, *Choreftria* **n. g.** shares several characters with three genera of the *Teredicola*-group. The genera of the

Teredicola-group were previously placed in the families Clausididae Embleton, 1901 or Clausiidae Giesbrecht, 1895 (see Wilson & Illg, 1955; Bocquet & Stock, 1958; Kim & Ho, 1991; Ho & Wardle, 1992; Kim, 2001). The taxonomic affinities of these genera are still controversial but they are often recognized as members of the family Clausidiidae. While the mandibles of the *Teredicola*-group genera, with three or four elements, are similar to that of Clausidiidae (see Humes, 1987; Kim & Stock, 1996; Ho & Kim, 2004; Kim, 2007, 2015; Moon & Kim, 2014; Hwang et al., 2016), that of Choreftria n. g. is reduced and bears only one apical serrated blade. Humes & Boxshall (1996) consider that the mandible is diagnostic at the family level in their revision of the lichomolgoid complex. Boxshall & O'Reilly (2015) recognized the similarity between the genus Gadilicola Boxshall & O'Reilly, 2015 and the genera of Teredicola-group but the independent family Gadilicolidae was established to accommodate the genus



Fig. 4 *Choreftria shiranui* **n. g., n. sp.**, adult female, holotype, NSMT-Cr 30590. A, left leg 1, anterior; B, right leg 2, anterior; C, right leg 3, anterior; D, right leg 4, anterior; E, left leg 5, ventral. Scale bars: A, 50 μm; B, 70 μm; C, D, 100 μm; E, 40 μm



because of it having a reduced mandible with an apical blade. This type of mandible is similar to that of *Choreftria* **n. g.**, and more like that of several genera of the families Clausiidae and Nereicolidae Claus, 1875, but other morphological features (e.g. body segmentation and other appendages, especially the mouth parts) are not similar between the new genus and these three families (see Kim et al., 2013; Boxshall & O'Reilly, 2015; present paper). Therefore, we herein establish a new monotypic family to accommodate *Choreftria* **n. g.** Just as Boxshall & O'Reilly (2015)

mentioned with Gadilicolidae, confirming the validity of the new family is required based on further phylogenetic research. Although the enlarged female's leg 4 of *Choreftria* **n. g.** is similar to that of *Telson* Pearse, 1952, there are many differences in other appendages between those two genera (see Causey, 1960; Ho, 1967). From sharing the maxilliped of female with a sigmoid bifurcate claw, *Telson* is considerable to be related to Bomolochidae Claus, 1875 and Makrostrotidae Huys et al., 2012 rather than Clausidiidae and its allied genera (see Huys et al.,



2012). The possession of two serrated mandibular blades of *Telson* also differs from *Choreftria* **n. g.**

While almost all of the genera that are possibly related to Choreftria n. g. (i.e. species of Teredicolagroup, Clausididae, Clausiidae, Gadilicolidae, and Nereicolidae) have been recorded from molluscs and polychaetes, C. shiranui n. g., n. sp. was found from a fish. There are several similar records among parasitic copepods of apparent host switching from invertebrate to vertebrate hosts. Members of the family Anthessiidae are known to be associated with molluscan hosts but Anthessius lophiomi Avdeev & Kazachenko, 1986 inhabits the angler fish, Lophiomus setigerus (Vahl) (Lophiiformes: Lophiidae) (Avdeev & Kazachenko, 1986). The family Macrochironidae is known to be associated with cnidarians but Paramacrochiron sewelli Reddiah, 1968, a member of the family, was recorded from the mackerel Trachurus trachurus (Linnaeus) (Perciformes: Carangidae) in addition to the scyphozoan jellyfish Lychnorhiza malayensis Stiasny (Rhizostomeae: Lychnorhizidae) (Reddiah, 1968) (Avdeev, 1975). Furthermore, in recent years, Pseudanthessius sp. which is a member of Pseudanthessiidae was described from the grouper Epinephelus sp. (Perciformes: Serranidae) while most of other known congeners have been found from various invertebrates (e.g. poriferans, turbellarians, gastropods, polychaetes, echinoids, crinoids, and ophiuroids) (e.g. Humes & Stock, 1973; Uyeno & Rain, 2021). This may be because of the extraordinary host change that occurred from invertebrates to fishes, and the finding of C. shiranui **n. g.**, **n. sp.** from the worm goby *Taenioides snyderi* might be a new example. It is probably not an accidental infection because the copepod was firmly attached to the host's anal fin by the antennae when the specimen was collected. Although C. shiranui n. g., n. sp. shows some similarities in the mouthparts with the copepods associated with marine invertebrates (i.e., with Teredicola-group, Clausididae, Clausiidae, Gadilicolidae, and Nereicolidae), it is difficult to define that C. shiranui n. g., n. sp. is commensal or parasitic at this stage. Since only one adult female of *C. shiranui* **n. g.**, **n. sp.** was found in this study, it is desirable that additional specimens will be found in the future to help understand its biology.

Choreftriidae n. fam.

LSID urn:lsid:zoobank.org:act:B5F24254-6CEC-4F5F-98BD-C1DDC5BAE3DF

New Japanese name: Oshiri-kajiri-mushi-ka. Diagnosis
As for the genus.

Type-genus: Choreftria **n. g.**, by original designation.

Acknowledgements I acknowledge Reo Koreeda (Kagoshima University) for providing the material, photograph, and biological information and Myles O'Reilly (Scottish Environment Protection Agency) for providing the valuable comments and kind corrections to improve the manuscript.

Author contributions DU has carried out all the procedures in the preparation of the manuscript.

Funding Part of this work was supported by the Japan Society for Promotion of Science (JSPS) KAKENHI Grant Number JP15H06517 and JP17K15304; "Establishment of Glocal Research and Education Network in the Amami Islands" project of Kagoshima University and "General and practical research for the restoration and creation of the natural environment in the Ariake Sea and the Yatsushiro Sea" project of Kumamoto University both adopted by the Ministry of Education, Culture, Sports, Science and Technology, Japan. Japan Society for Promotion Science (JSPS),JP15H06517,DAISUKE UYENO JP17K15304,DAISUKE UYENO .the Ministry Education, Culture, Sports, Science and Technology, Japan.

Declarations

Conflict of interest The author has no competing interests to declare.

Data availability See Material Examined section.

References

Avdeev, G. V. (1975). The first case of discovery of Lichomolgidae member (Copepoda, Cyclopoida) on a vertebrate animal from the region of Sri Lanka. *Parazitologiya*, 9, 202–204 (**Russian with English summary**).

Avdeev, G. V., & Kazachenko, V. N. (1986). Parasitic copepods from fishes of the genus *Lophiomus* Gill in the Pacific. *Crustaceana*, 50, 53–67.

Bocquet, C., & Stock, J. H. (1958). Copépodes parasites d'invertébrés des côtes de la Manche. IV. Sur les trois genres synonymes de copépodes cyclopoïdes, *Leptinogaster*



- Pelseneer, Strongylopleura Pelseneer et Myocheres Wilson (Clausidiidae). Archives de Zoologie Expérimentale et Générale 96, notes et revue 2, 71–89.
- Boxshall, G. A., & Halsey, S. H. (2004). An introduction to copepod diversity. London: The Ray Society, 966 pp.
- Boxshall, G. A., & O'Reilly, M. (2015). The first parasitic copepod from a scaphopod mollusc host. *Systematic Parasitology*, *90*, 113–124.
- Causey, D. (1960). Parasitic copepoda from Mexican coastal fishes. Bulletin of Marine Science of the Gulf and Caribbean, 10, 323–337.
- Froese, R., & Pauly, D. (Eds.) (2021). *FishBase*. World Wide Web electronic publication. www.fishbase.org, version (06/2021).
- Ho, J. S. (1967). Cyclopoid copepods of the genus *Telson* parasitic on Uranoscopid fishes in the Gulf of Mexico. *Journal of Parasitology*, 53, 852–858.
- Ho, J. S., & Kim, I. H. (2004). A new genus of the Clausidiidae (Copepoda: Poecilostomatoida) associated with a polychaete from Korea, with discussion of the taxonomic status of *Hersiliodes Canu*, 1888. *Proceedings of the Biological* Society of Washington, 117, 95–105.
- Ho, J. S., & Wardle, W. J. (1992). Pholadicola intestinalis, new genus and species, a clausidiid copepod parasitic in a deepburrowing clam from Texas. Bulletin of Marine Science, 51, 37–44.
- Humes, A. G. (1987). Copepods from deep-sea hydrothermal vents. *Bulletin of Marine Science*, 41, 645–788.
- Humes, A. G., & Boxshall, G. A. (1996). A revision of the lichomolgoid complex (Copepoda: Poecilostomatoida), with the recognition of six new families. *Journal of Natural History*, 30, 175–227.
- Humes, A. G., & Cressey, R. F. (1958). Copepod parasites of mollusks in West Africa. *Bulletin de l'Institut Français d'Afrique Noire (A)*, 20, 921–942.
- Humes, A. G., & Gooding, R. U. (1964). A method for studying the external anatomy of copepods. *Crustaceana*, 6, 238–240.
- Humes, A.G., & Stock, J. H. (1973). A revision of the family Lichomolgidae Kossman, 1877, cyclopoid copepods mainly associated with marine invertebrates. *Smithsonian Contributions to Zoology*, 127, 1–368.
- Humes, A. G., & Turner, R. D. (1972). Teredicola typicus C.B. Wilson, 1942 (Copepoda, Cyclopoida) from shipworms in Australia, New Zealand, and Japan. Australian Journal of Marine and Freshwater Research, 23, 63–72.
- Huys, R., Fatih, F., Ohtsuka, S., & Llewellyn-Hughes, J. (2012). Evolution of the bomolochiform superfamily complex (Copepoda: Cyclopoida): New insights from ssrDNA and morphology, and origin of umazuracolids from polychaeteinfesting ancestors rejected. *International Journal for Parasitology*, 42, 71–92.

- Hwang, H. S., Lee, J. M., & Kim, I. H. (2016). Two new species of Clausidiidae (Copepoda, Poecilostomatoida) from Korea. Animal Systematics Evolution and Diversity, 32, 93–104.
- Kim, I. H. (2001). Foliomolgus cucullus, a new genus and species of Clausidiidae (Crustacea: Copepoda: Poecilostomatoida) associated with a polychaete in Korea. Proceedings of the Biological Society of Washington, 114, 660–666.
- Kim, I. H. (2007). Giardella ricoensis n. sp. from the West Indies and the proposal of the new genus Goodingius (Copepoda: Cyclopoida: Clausidiidae). Korean Journal of Systematic Zoology, 23, 127–133.
- Kim, I. H. (2009). Poecilostome copepods (Crustacea: Cyclopoida) associated with marine invertebrates from tropical waters. Korean Journal of Systematic Zoology, Special Issue No.7. 7, 1–90.
- Kim, I. H. (2015). Hamaticyclops ahni, a new genus and species of the family Clausidiidae (Crustacea, Copepoda, Poecilostomatoida) from California, United States. Animal Systematics Evolution and Diversity, 31, 304–310.
- Kim, I. H., & Ho, J. S. (1991). Copepod parasites of commercial bivalves from Korea. I. Two new poecilostomatoid species from *Solen grandis* Dunker in the Yellow Sea. *Korean Journal of Systematic Zoology*, 7, 1–11.
- Kim, I. H., Sikorski, A., O'Reilly, M. G., & Boxshall, G. A. (2013). Copepods associated with polychaete worms in European seas. *Zootaxa*, 3651, 1–62.
- Kim, I. H., & Stock, J. H. (1996). A new species of Clausidiidae (Copepoda, Poecilostomatoida) associated with the bivalve Ruditapes philippinarum in Korea. Cahiers de Biologie Marine, 37, 1–6.
- Moon, S. Y., & Kim, I. H. (2014). Description of *Conchyliurus dispar* n. sp. (Copepoda, Cyclopoida, Clausidiidae) associated with the bivalve *Barnea manilensis* (Philippi) from the Yellow Sea with a discussion of the male morphotypes in the genus. *Zootaxa*, 3760, 471–478.
- Uyeno, D., & Rain, E. (2021). A new species of *Pseudanthessius* (Copepoda: Cyclopoida: Pseudanthessiidae) on *Epine-phelus* sp. (Actinopterygii: Perciformes: Serranidae) from Chuuk Lagoon, Federated States of Micronesia. *Species Diversity*, 26, 249–254.
- Wilson, M. S. (1957). Redescription of *Teredicola typica* C.B. Wilson (Crustacea: Copepoda). *Pacific Science*, 11, 265–274.
- Wilson, M. S., & Illg, P. L. (1955). The family Clausiidae (Copepoda, Cyclopoida). Proceedings of the Biological Society of Washington, 68, 129–142.
- **Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Terms and Conditions

Springer Nature journal content, brought to you courtesy of Springer Nature Customer Service Center GmbH ("Springer Nature"). Springer Nature supports a reasonable amount of sharing of research papers by authors, subscribers and authorised users ("Users"), for small-scale personal, non-commercial use provided that all copyright, trade and service marks and other proprietary notices are maintained. By accessing, sharing, receiving or otherwise using the Springer Nature journal content you agree to these terms of use ("Terms"). For these purposes, Springer Nature considers academic use (by researchers and students) to be non-commercial.

These Terms are supplementary and will apply in addition to any applicable website terms and conditions, a relevant site licence or a personal subscription. These Terms will prevail over any conflict or ambiguity with regards to the relevant terms, a site licence or a personal subscription (to the extent of the conflict or ambiguity only). For Creative Commons-licensed articles, the terms of the Creative Commons license used will apply.

We collect and use personal data to provide access to the Springer Nature journal content. We may also use these personal data internally within ResearchGate and Springer Nature and as agreed share it, in an anonymised way, for purposes of tracking, analysis and reporting. We will not otherwise disclose your personal data outside the ResearchGate or the Springer Nature group of companies unless we have your permission as detailed in the Privacy Policy.

While Users may use the Springer Nature journal content for small scale, personal non-commercial use, it is important to note that Users may not:

- 1. use such content for the purpose of providing other users with access on a regular or large scale basis or as a means to circumvent access control;
- 2. use such content where to do so would be considered a criminal or statutory offence in any jurisdiction, or gives rise to civil liability, or is otherwise unlawful;
- 3. falsely or misleadingly imply or suggest endorsement, approval, sponsorship, or association unless explicitly agreed to by Springer Nature in writing;
- 4. use bots or other automated methods to access the content or redirect messages
- 5. override any security feature or exclusionary protocol; or
- 6. share the content in order to create substitute for Springer Nature products or services or a systematic database of Springer Nature journal content.

In line with the restriction against commercial use, Springer Nature does not permit the creation of a product or service that creates revenue, royalties, rent or income from our content or its inclusion as part of a paid for service or for other commercial gain. Springer Nature journal content cannot be used for inter-library loans and librarians may not upload Springer Nature journal content on a large scale into their, or any other, institutional repository.

These terms of use are reviewed regularly and may be amended at any time. Springer Nature is not obligated to publish any information or content on this website and may remove it or features or functionality at our sole discretion, at any time with or without notice. Springer Nature may revoke this licence to you at any time and remove access to any copies of the Springer Nature journal content which have been saved.

To the fullest extent permitted by law, Springer Nature makes no warranties, representations or guarantees to Users, either express or implied with respect to the Springer nature journal content and all parties disclaim and waive any implied warranties or warranties imposed by law, including merchantability or fitness for any particular purpose.

Please note that these rights do not automatically extend to content, data or other material published by Springer Nature that may be licensed from third parties.

If you would like to use or distribute our Springer Nature journal content to a wider audience or on a regular basis or in any other manner not expressly permitted by these Terms, please contact Springer Nature at

onlineservice@springernature.com